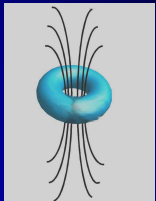


# Biomagnetism:

## Detection and Imaging Weak Magnetic Fields from the Human Body



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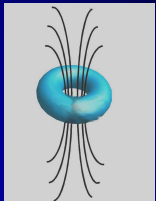
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# Senior Scientific, LLC:

A private business with NIH SBIR funding.

Primary Mission: Disease detection using magnetic nanotechnology.

PI: Edward R. Flynn, PhD Physicist



Senior Scientific

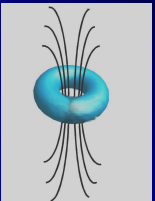


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# Outline of Talk

## Detection and Imaging of Weak Biomagnetic Fields

- 1) SQUID Sensors
- 2) Experimental Apparatus
- 3) Imaging the Brain
- 4) Imaging the Mind
- 5) Detection and Imaging Disease

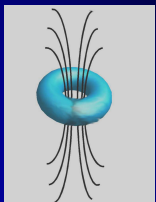


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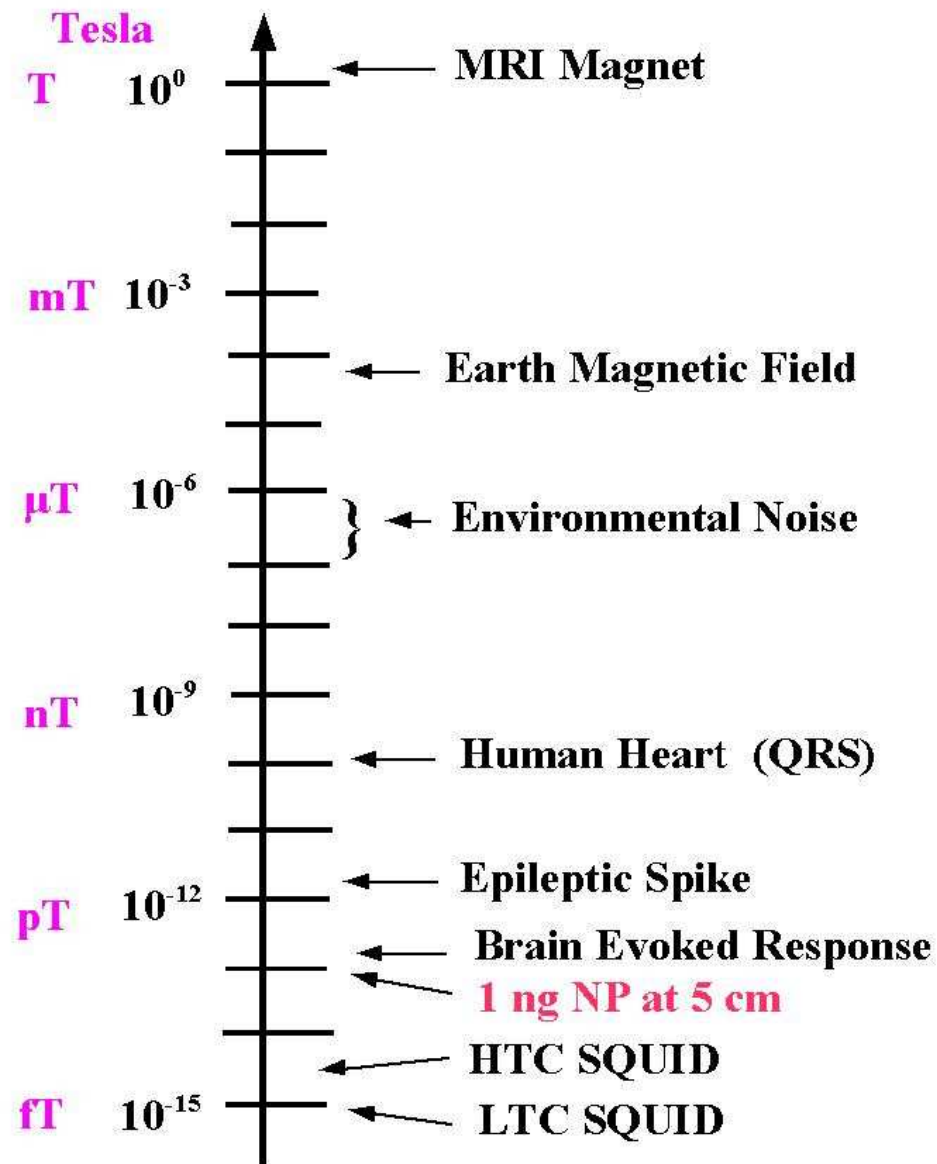
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# Magnetic Field Strengths of various sources and Sensor Sensitivities



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## Magnetic Field Strength



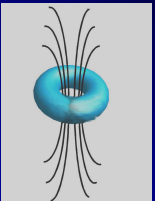
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# MEG

## Magnetoencephalography

Measurement of the natural magnetic fields  
From the brain arising from neuronal currents

A: Instrumentation



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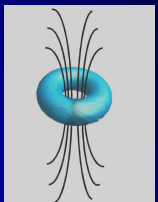
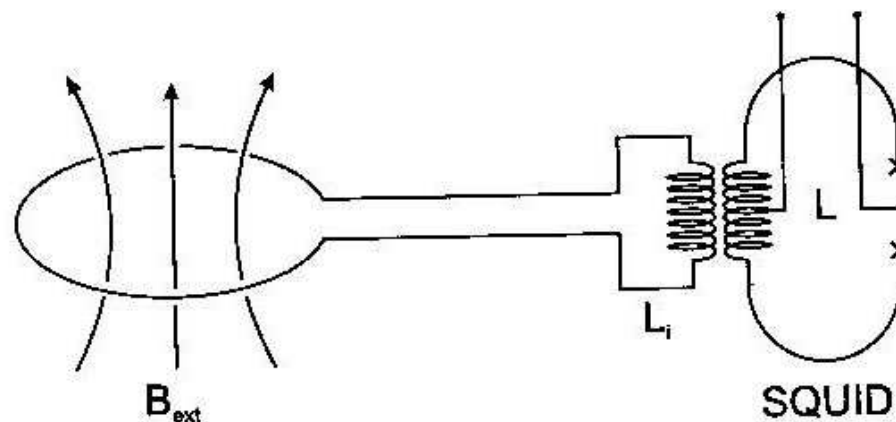
# What is a SQUID?

Superconducting Quantum Interference Device (SQUID)

Quantum Mechanical (Josephson) Tunneling in a Superconductor

Sensor must have no resistive noise so superconducting material is used.

• LTC SQUIDs are sensitive down to  $\sim 3$  fT

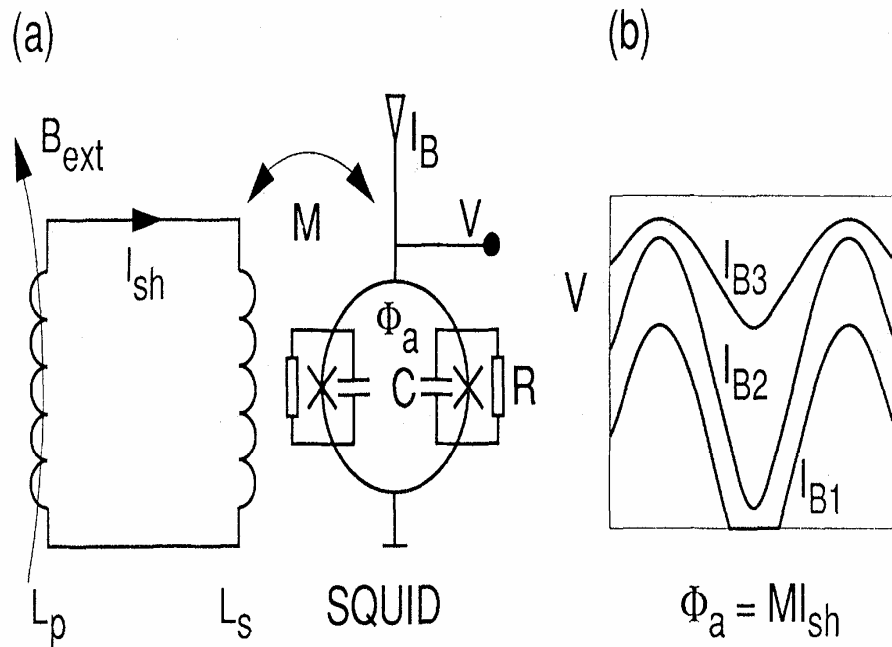


# Basics of SQUID Operation

$$I = I_c \sin \theta ,$$

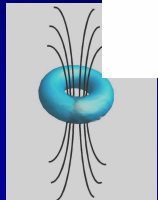
$$\frac{\partial \theta}{\partial t} = 2\pi V \frac{2e}{h} = \frac{2\pi V}{\Phi_0}$$

Josephson Equations:  $\theta$  is phase across junction,  $I$ =current,  $V$ = junction voltage,  $\Phi_a = h/2e=2.07$  fWb (flux quanta)

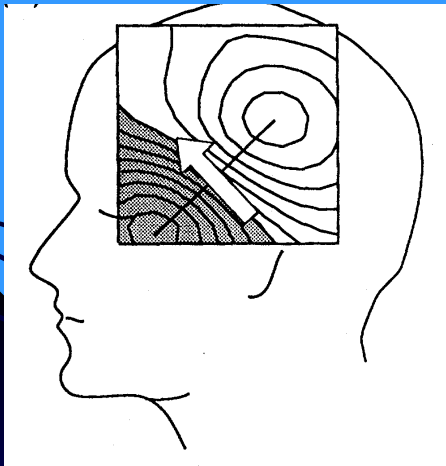
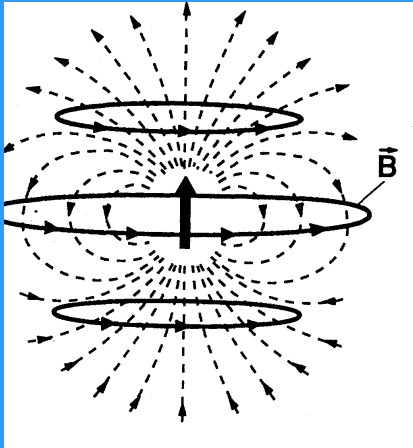


(a) Schematic diagram of DC SQUID,  $L_s$  is superconducting flux transformer

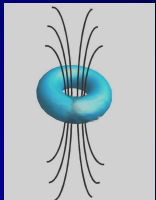
(b) Voltage across SQUID depends on bias  $I$ , and is periodic function of the incident flux  $\Phi_a$ .



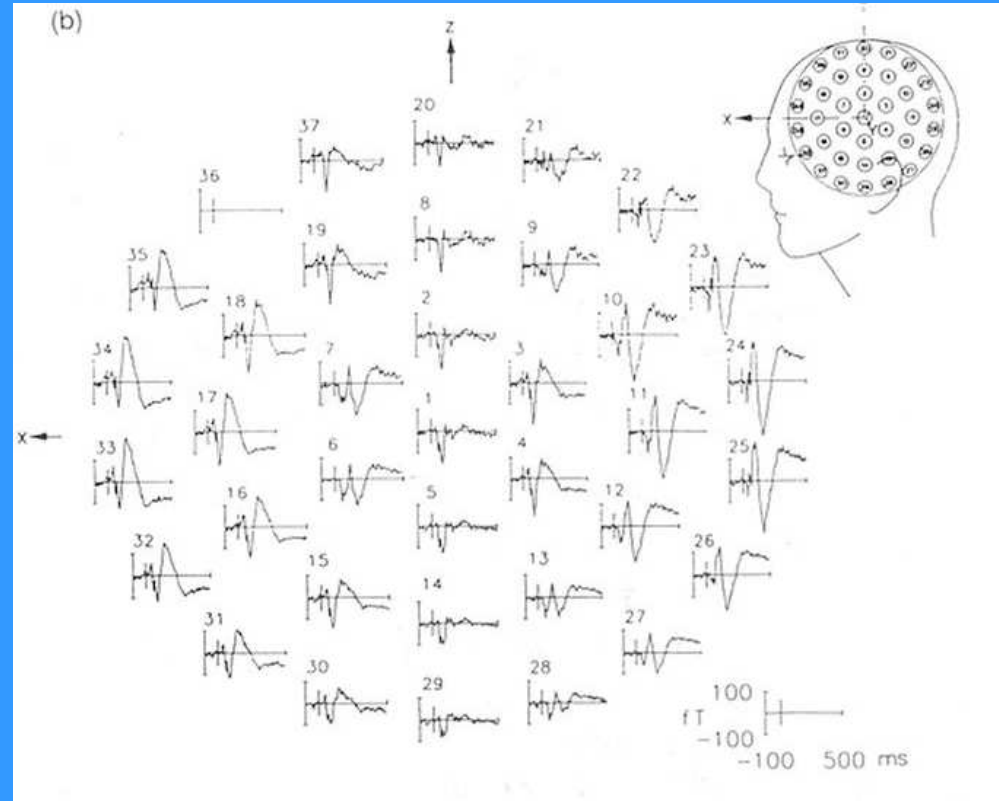
# Principles of MEG



Current Dipole



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Field amplitude vs time for a sensor array - Evoked Response

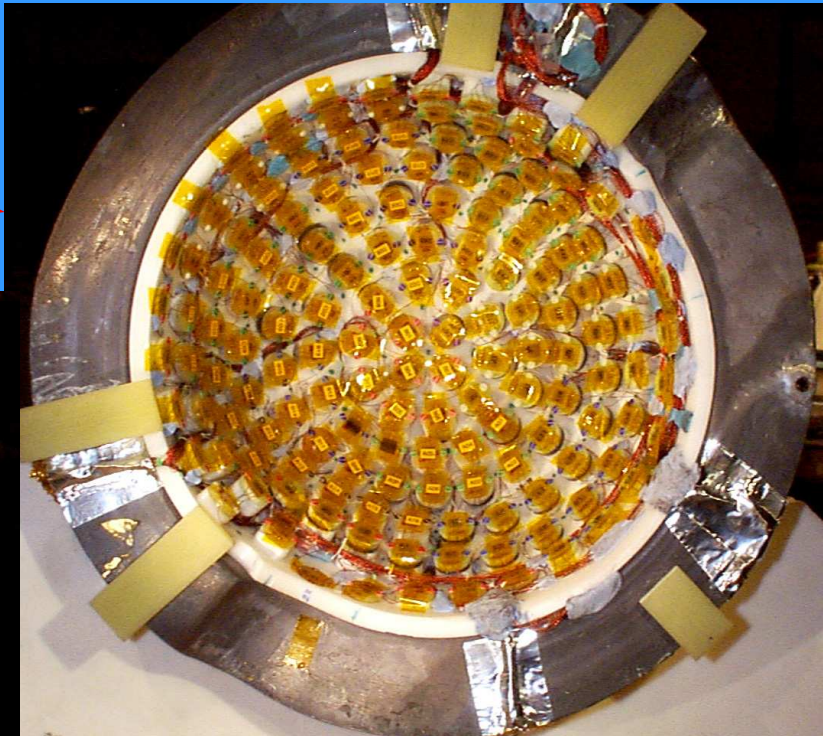
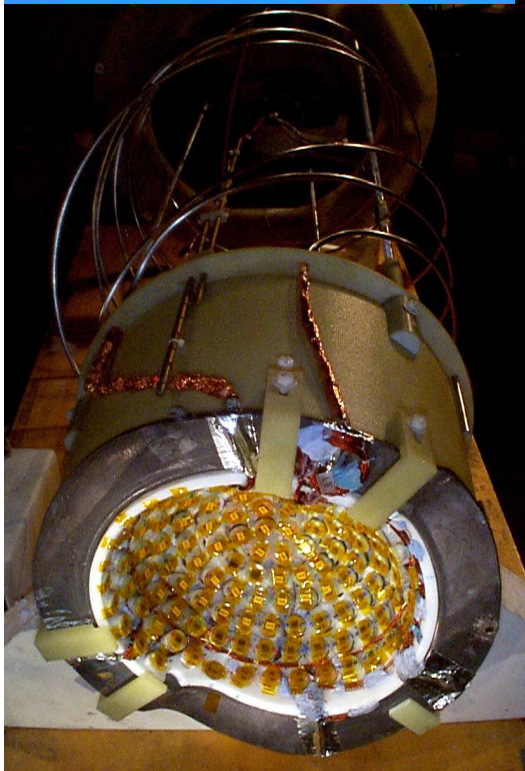
Matti Hämäläinen, Ritta Hari, Risto J. Ilmoniemi, Jukka Knutila, and Olli V. Lounasmaa  
to noninvasive studies of the working human brain  
Magnetoencephalography—theory, instrumentation, and applications

Reviews of Modern Physics, Vol. 65, No. 2, April 1993



# The SISG MEG System

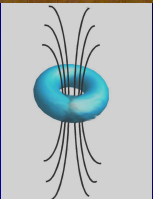
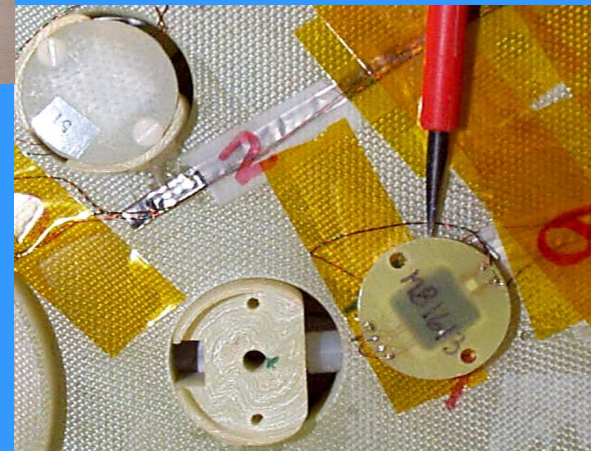
Superconducting  
"helmet" made of  
a thin lead sheet.



155 ch SQUID  
array installed  
inside  
superconducting  
imaging surface  
"helmet"

Integrated  
SQUID sensors  
and pickup coils

A large array sensor for MEG  
based on the superconducting  
imaging surface gradiometer  
concept.

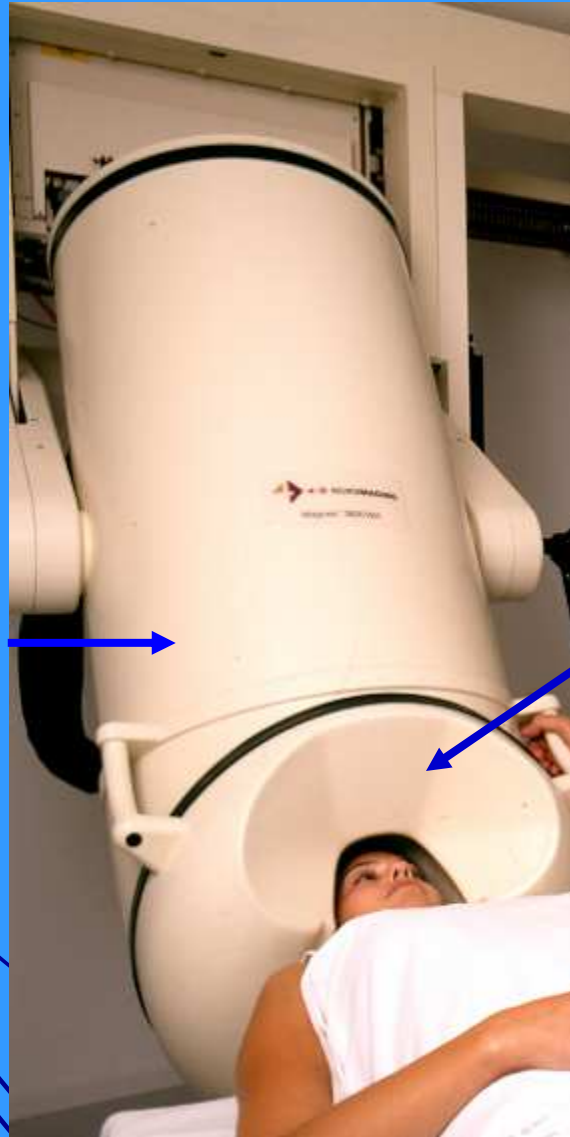


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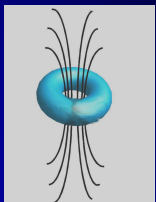


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The Dewar  
Liquid helium



- 248 axial gradiometers (low noise)
- 1 kHz sampling rate



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The MEG instrument at the Minneapolis Domenici Center  
(Magnes 3600WH, 4-D Neuroimaging, San Diego, CA)



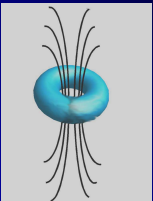
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# MEG

## Magnetoencephalography

Imaging the Brain

SpatioTemporal Analysis of  
Sensor Magnetic Fields to  
Image Brain Sources using  
EM Inverse Solutions



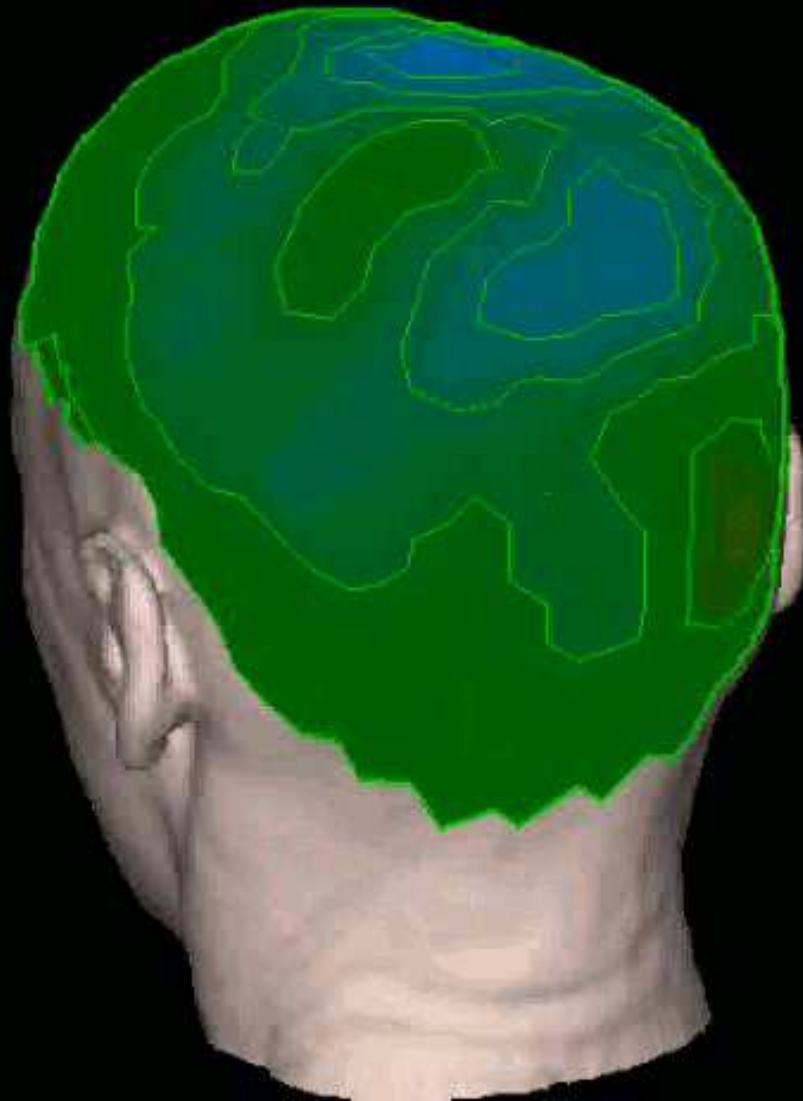
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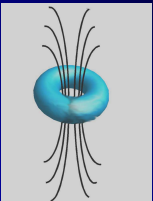
# SQUIDS Measure Both Space and Time

milliseconds: 50



Magnetic field contour lines, plotted here as a function of time, are used to determine neural sources.

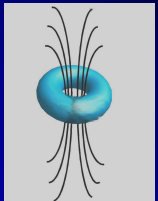
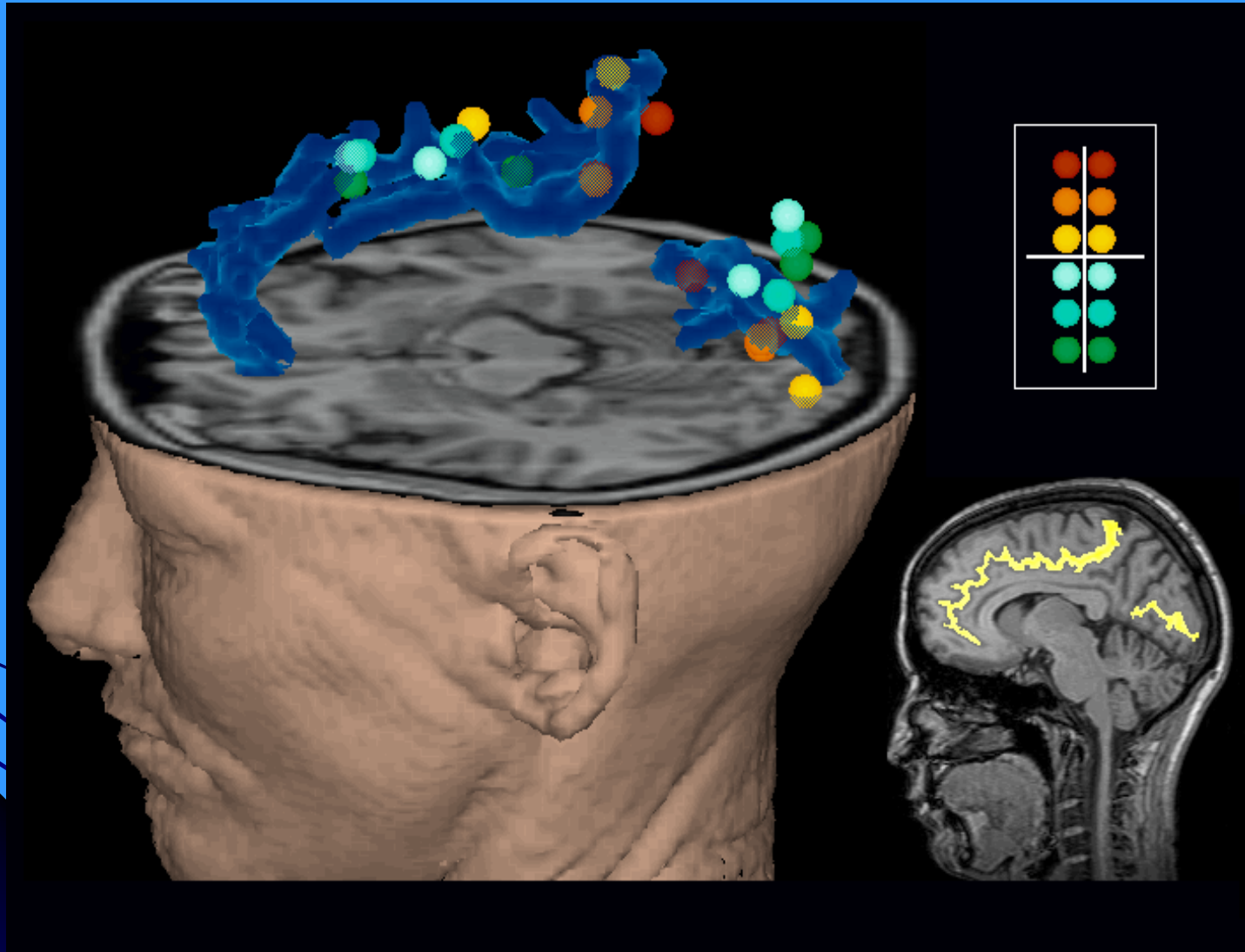
Response of the brain to a visual stimulus.



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# Finding the Sources with a Spatial-Temporal algorithm



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Graphics by  
Ranken LANL

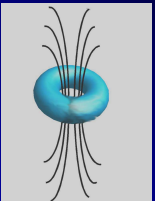


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# MEG

## Magnetoencephalography

Imaging the Mind  
Examine correlations between  
magnetic field magnitude and  
time in sensor space  
(No inverse problem)



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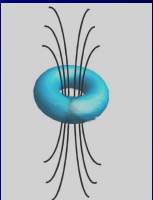
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# Data Analysis - 1

Analyses are performed to estimate quantitatively the synchronous (i.e. zero-lag) interactions between signals from pairs of sensors to assess dynamic brain function.

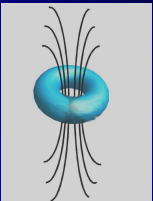
- Step 1: Calculate all pairwise zero-lag cross-correlations
- Step 2: Calculate the partial zero-lag cross-correlations within the 248-sensor network

Langheim, F.J.P., Leuthold, A.C. and Georgopoulos, A.P. (2006) Synchronous dynamic brain networks revealed by magnetoencephalography (MEG) Proceedings of the National Academy of Sciences USA 103: 455-459.



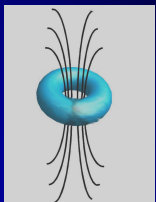
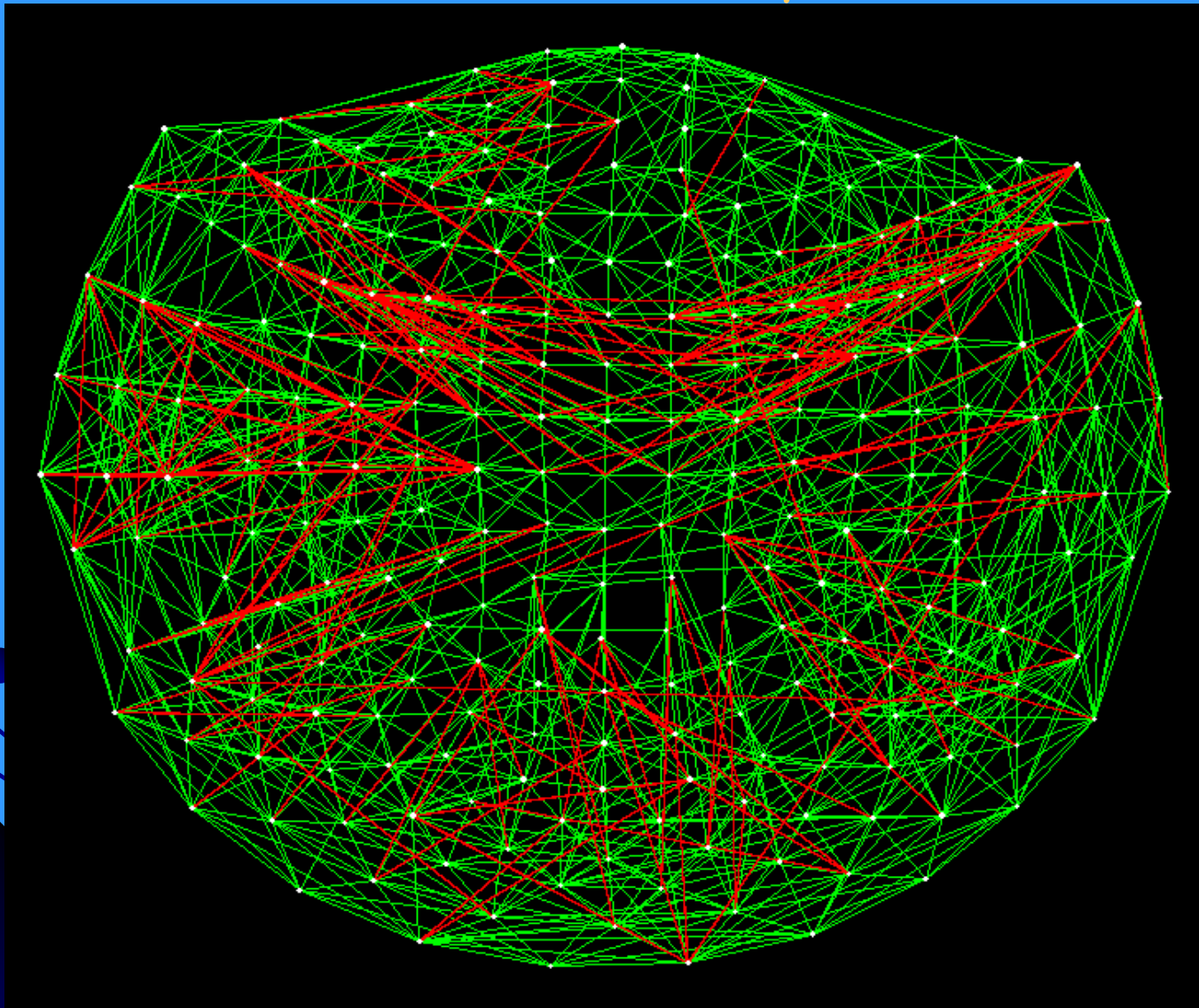
## Data Analysis - 2

- The MEG time series are “prewhitened” by fitting an ARIMA (AutoRegressive Integrative Moving Average) Box-Jenkins model and taking the residuals
- This procedure yields practically stationary series from which CCF is estimated





# Zero-lag (1-ms synchronous) Partial Correlations Of Prewhitened (stationary) MEG Time Series



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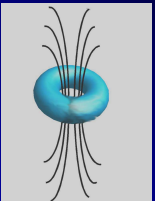
Langheim et al., PNAS, 2006



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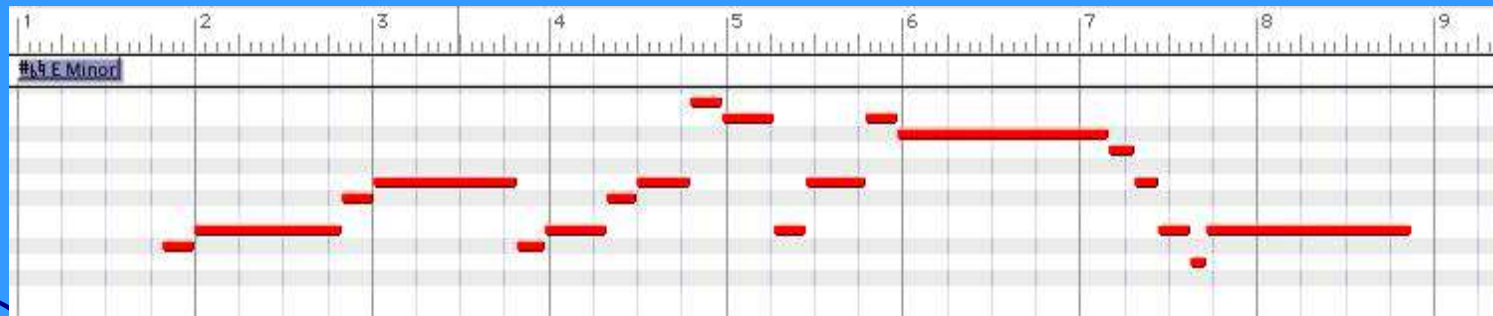
# Predictions from raw MEG signals: Music Perception

- Subjects listened to a musical piece while MEG was recorded
- Single trials analyzed using multivariate regression of MEG data on MIDI notes of the piece
- Predicting MIDI notes listened to



# Music Stimulus A

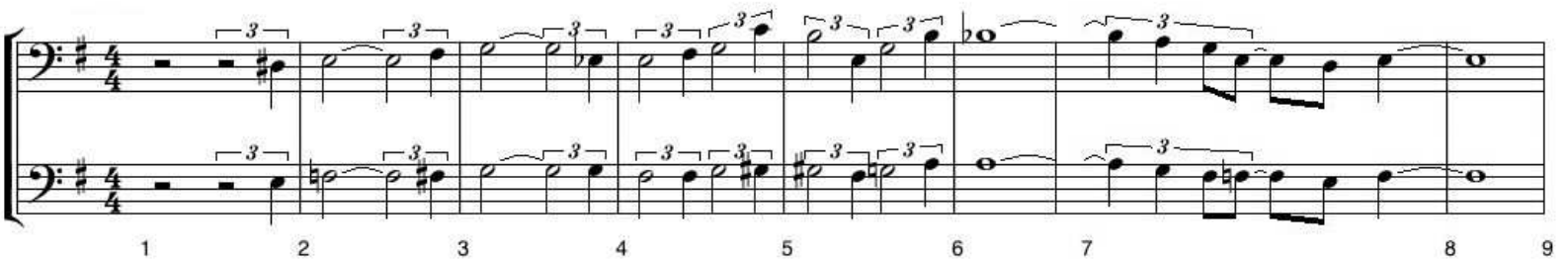
PPanther\_Stimulus



# Music Prediction A

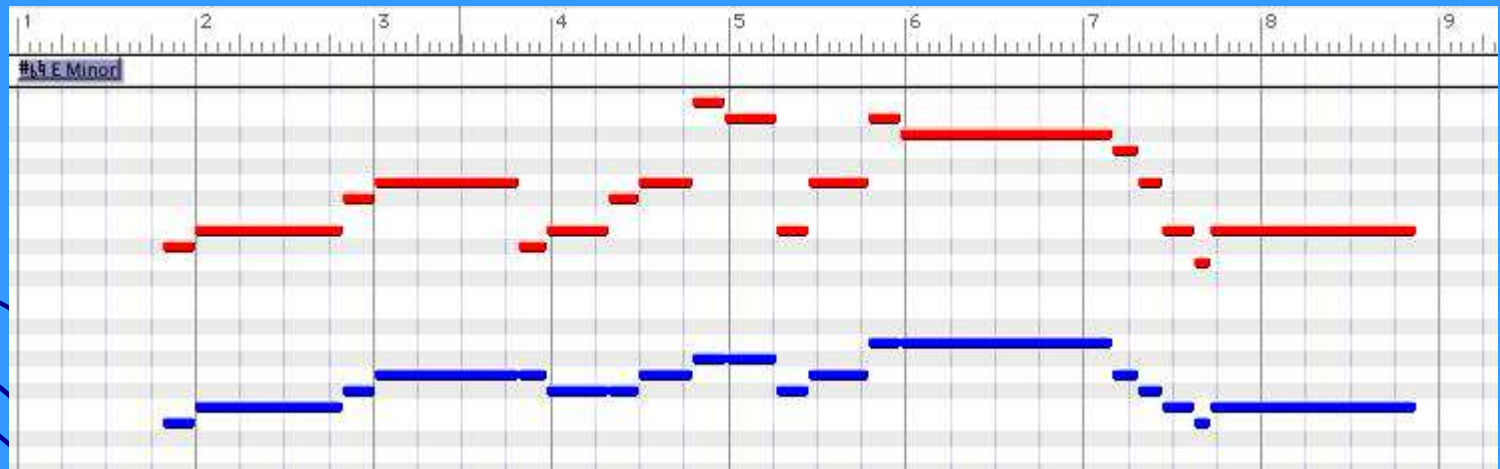
PPanther\_Stimulus

PPanther\_Prediction



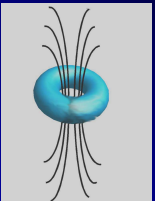
1 2 3 4 5 6 7 8 9

The image displays two staves of musical notation in bass clef, 4/4 time, with a key signature of one sharp (F#). The top staff, labeled 'PPanther\_Stimulus', contains a sequence of notes with triplets and slurs. The bottom staff, labeled 'PPanther\_Prediction', shows a similar sequence of notes, also with triplets and slurs. The notation is aligned with a horizontal axis numbered 1 through 9.



# Assessment of Dynamic Brain Function: Synchronous Neural Networks

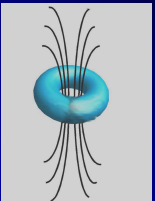
- Examine correlations between magnetic field magnitude and time in sensor space
- All possible zero-lag partial cross-correlations between 248 sensors (= 30,628)
- Positive or negative
- 1-ms temporal resolution = true synchronicity
- Simple fixation - look at a dot for 45 - 60 sec



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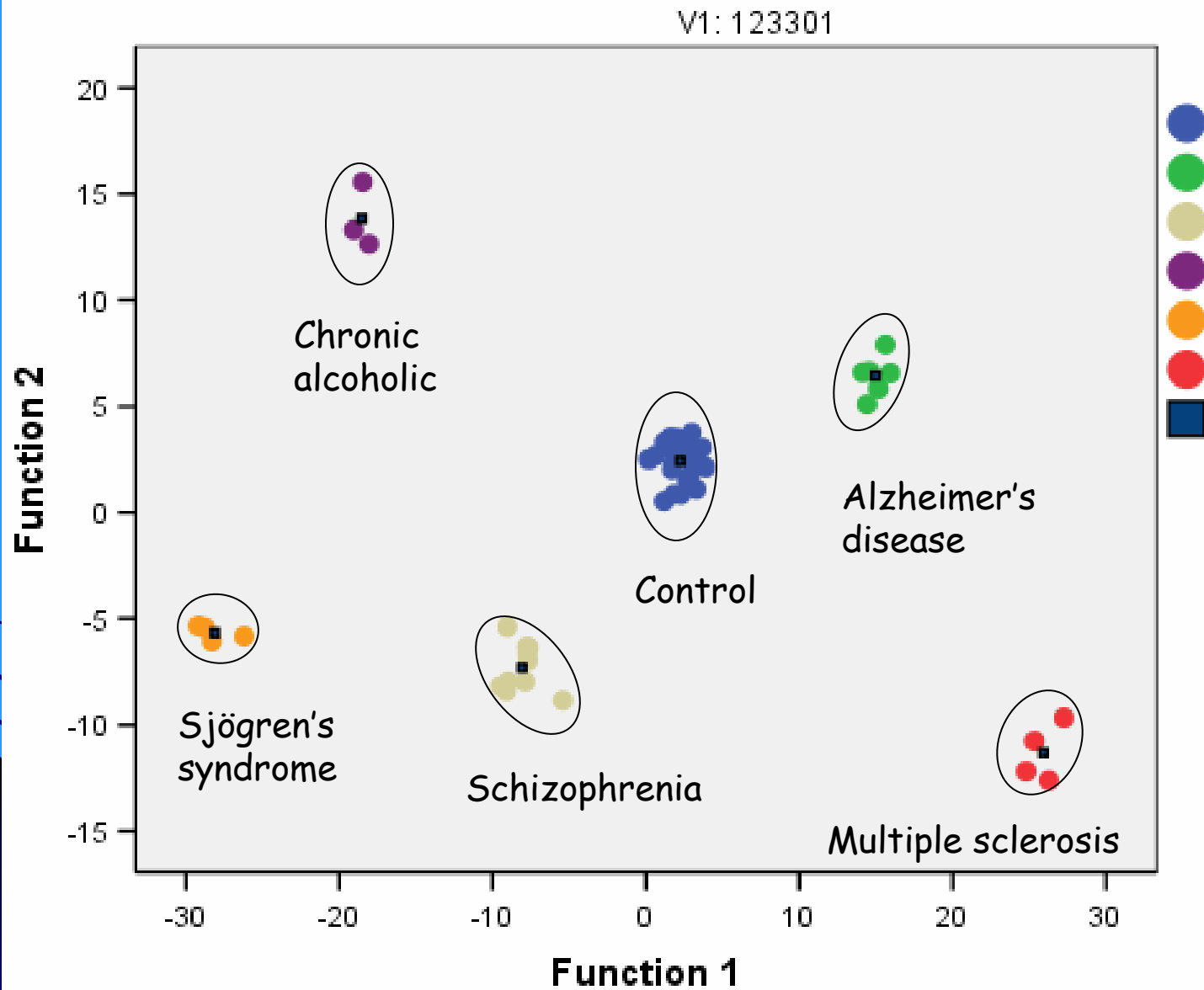
# Discriminant classification analysis

- Linear discriminant analysis
- Robust, cross-validated leave-one-out method
- 100% correct classification of 52 subjects to one of 6 groups (healthy control, Alzheimer's Disease, schizophrenia, chronic alcoholic, multiple sclerosis, Sjögren's syndrome) using as few as 10 zero-lag cross-correlations as predictors!
- Such sets are found in numbers far in excess of those expected by chance

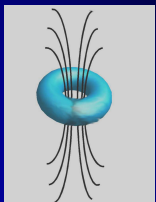




# Canonical Discriminant Functions



40 predictors



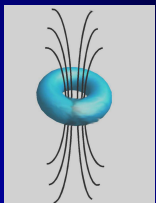
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# Superparamagnetic Particles and the Detection and Imaging of Disease using Magnetic Sensors

## Superparamagnetism



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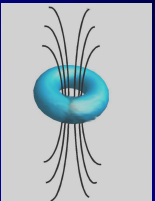
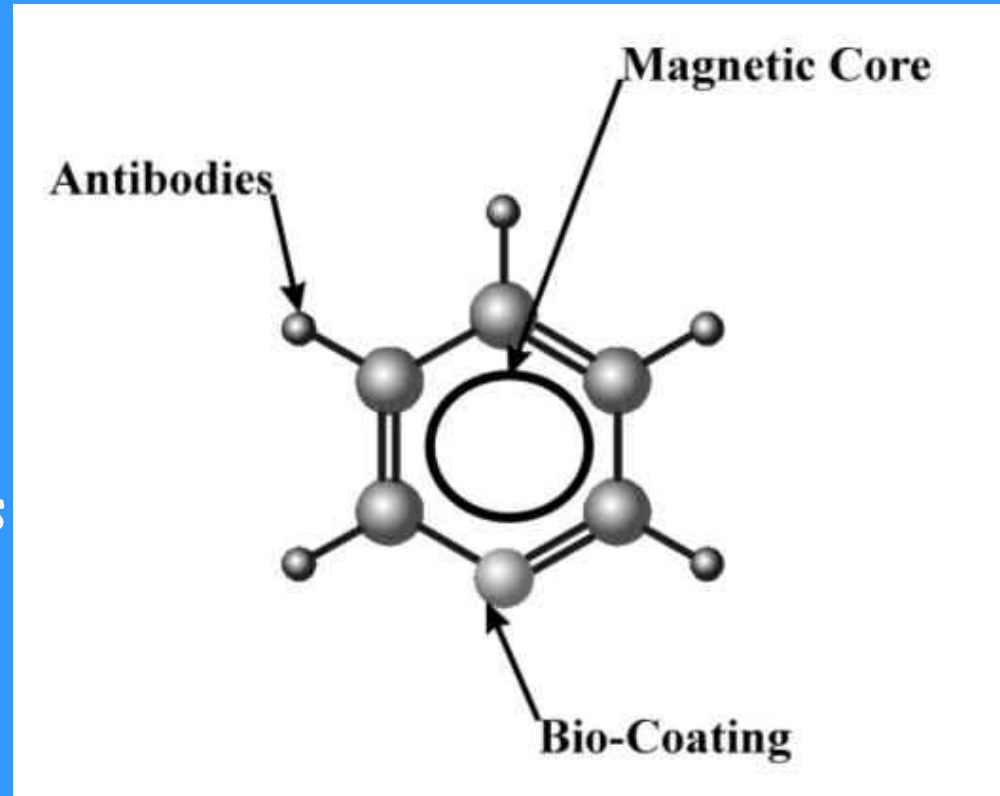


# Magnetic Nanoparticle with antibody attached

Typical magnetic core diameter is 20 -30 nm.

Typical bio-coatings are:  
Carboxyl, starch,  
streptavidin, PEG

Antibodies are specific markers for various types of cells; e.g., T-cells, various types of cancer cells, etc.



# Superparamagnetism

Iron-oxide nanoparticles <100 nm diameter

Particles consist of a single magnetic domain.

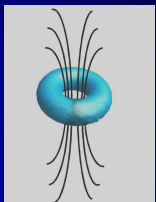
All internal atomic magnetic moments are aligned (homogeneous magnetization)

Free particles randomize quickly by Brownian Motion

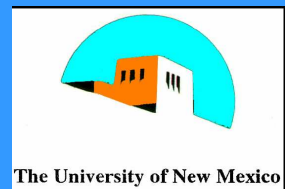
Bound particles decay by Néel Mechanism

Particles exhibit large magnetic moments when magnetized

Particles behave as paramagnetic when not magnetized (they do not agglomerate)



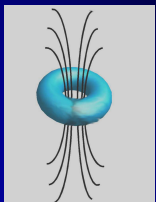
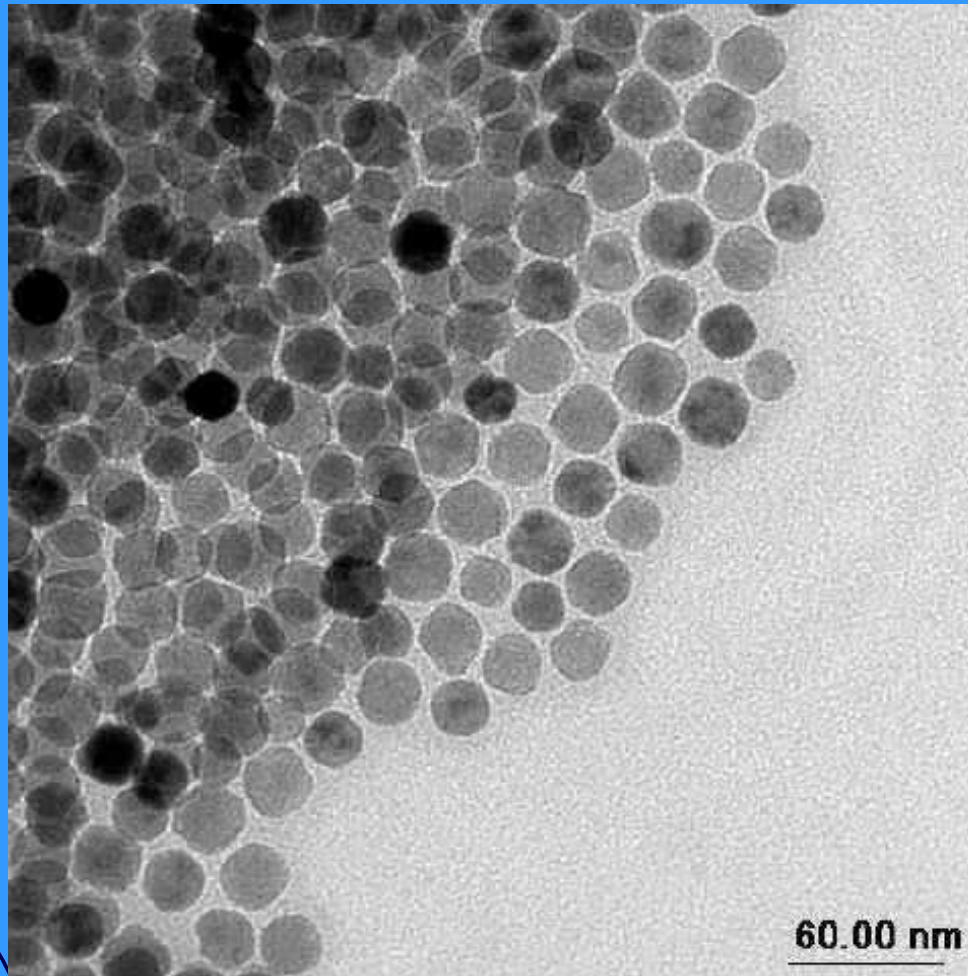
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# Scanning Electron Microscope View of Nanoparticles

Monodisperse magnetite 20 nm diameter, made at Center for Integrative Nanotechnology at Sandia National Laboratory (Dale Huber)



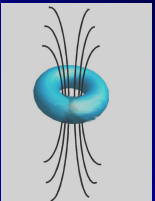
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# Nanoparticles form a Magnetic Dipole when a magnetizing pulse is applied

- An induced collective dipole moment  $\mu(t)$  is the result of alignment of a collection of  $N$  particles each with a moment  $\mu_p$  by an external pulsed field  $B$  for a duration  $t_0$ .
- $\mu(t)$  decays as the individual particle orientations relax, this is called the remanence time.



# The interaction of a nanoparticle of magnetic moment $\mu$ with a magnetic field

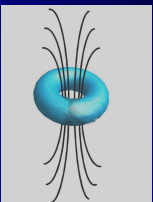
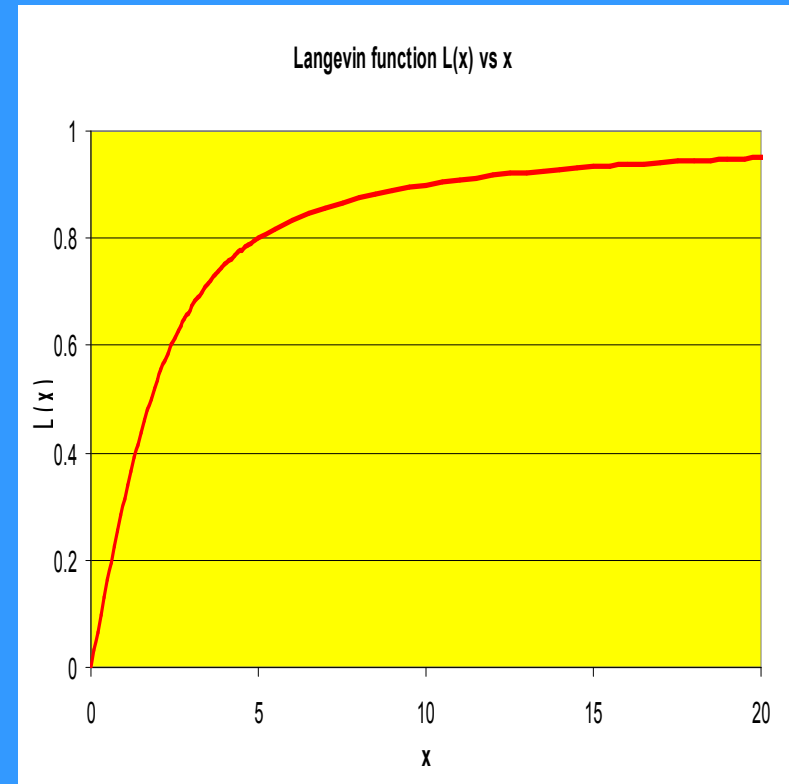
$$U = -\vec{\mu} \cdot \vec{B}$$

The average value of the cosine of the angle between is

$$\overline{\cos \theta} = \int e^{-U/kT} \cos \theta d\Omega / \int e^{-U/kT} d\Omega$$

The Langevin function,  $L(x)$  gives the average value of  $\cos \theta$  where  $x = \mu B/kT$

$$L(x) = \coth(x) - 1/x$$



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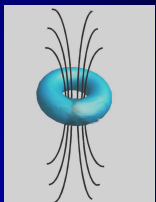
# Dipole formed by the magnetic pulse and its decay

Each nanoparticle of radius  $r$  is aligned by the field of pulsed Helmholtz coils to form an initial moment determined by the Langevin function and the Néel relaxation time:

$$\mu_0(r, t_0, B) = \mu_p L(x) [1 - \exp(-t_0 / \tau(r, B))]$$

The decaying dipole seen by a SQUID is the sum of all the aligned nanoparticle moments as they randomize when the field is quenched.

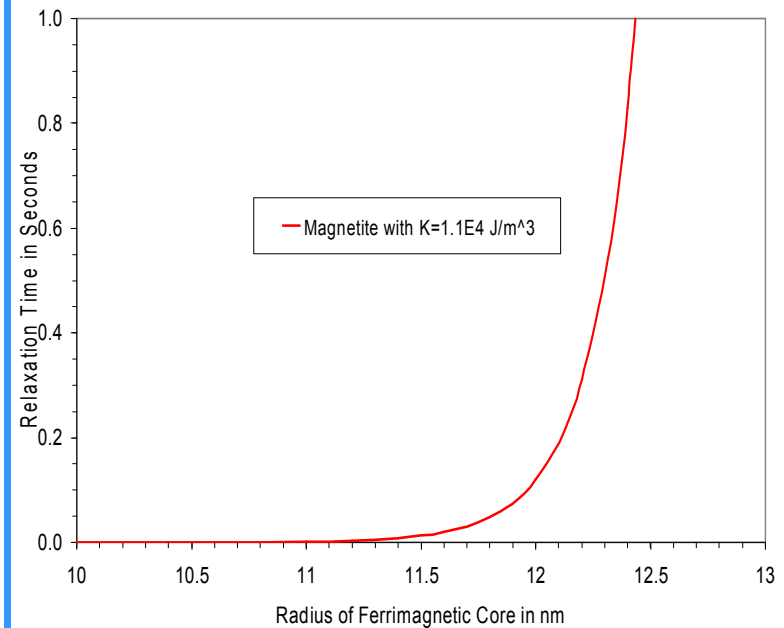
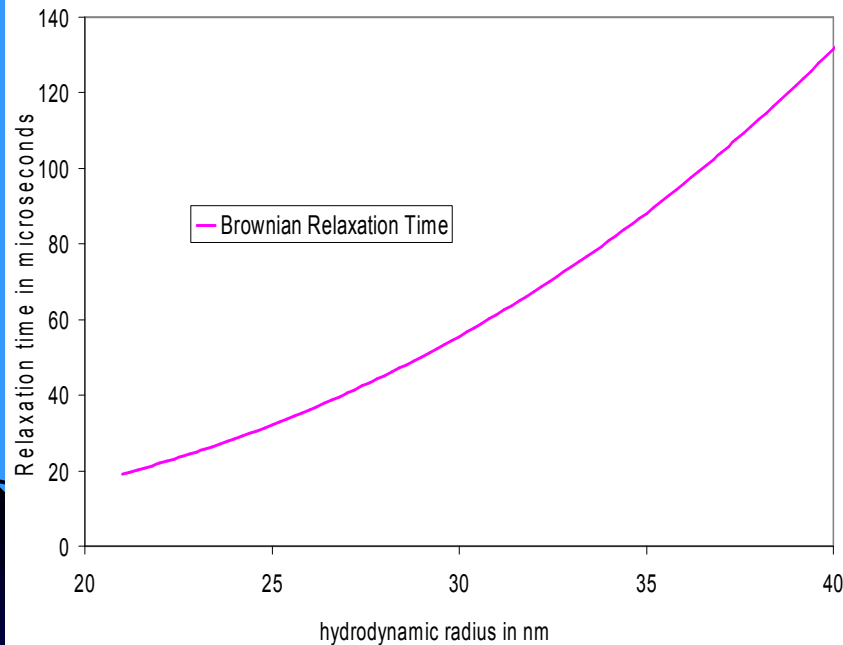
$$\mu(t, t_0, B) = N \int_0^\infty dr P(r) \mu_0(r, t_0, B) \exp(-t / \tau(r, 0))$$



# Brownian vs Néel Relaxation Times

Free Particles

Bound Particles



Brownian

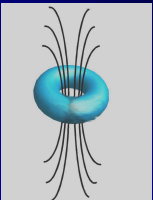
Néel

$$\tau_{Brownian} = 4\pi\eta R^3 / kT$$

$$\tau_{Neel} = \tau_0 \exp(KV / kT)$$

For polydisperse nanoparticles

$$B(t) = a_0 + a_1 * \ln(1+b/t)$$



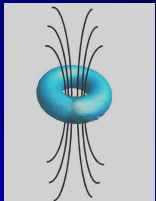
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# Superparamagnetic Particles and the Detection and Imaging of Disease

## Measuring the Remanence Fields



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# 7-Channel SQUID Measuring Chamber

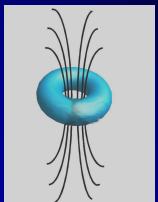
SQUIDs

Cell  
Source

Helmholtz  
Coils

Non-Magnetic  
Stage

Multi-channel system permits  
vector moment measurements

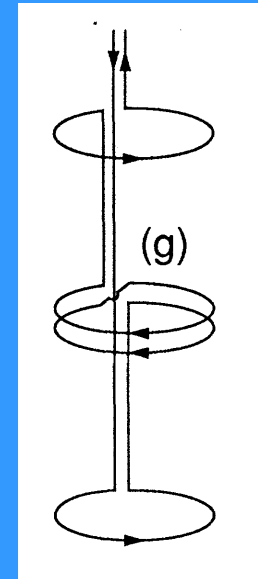
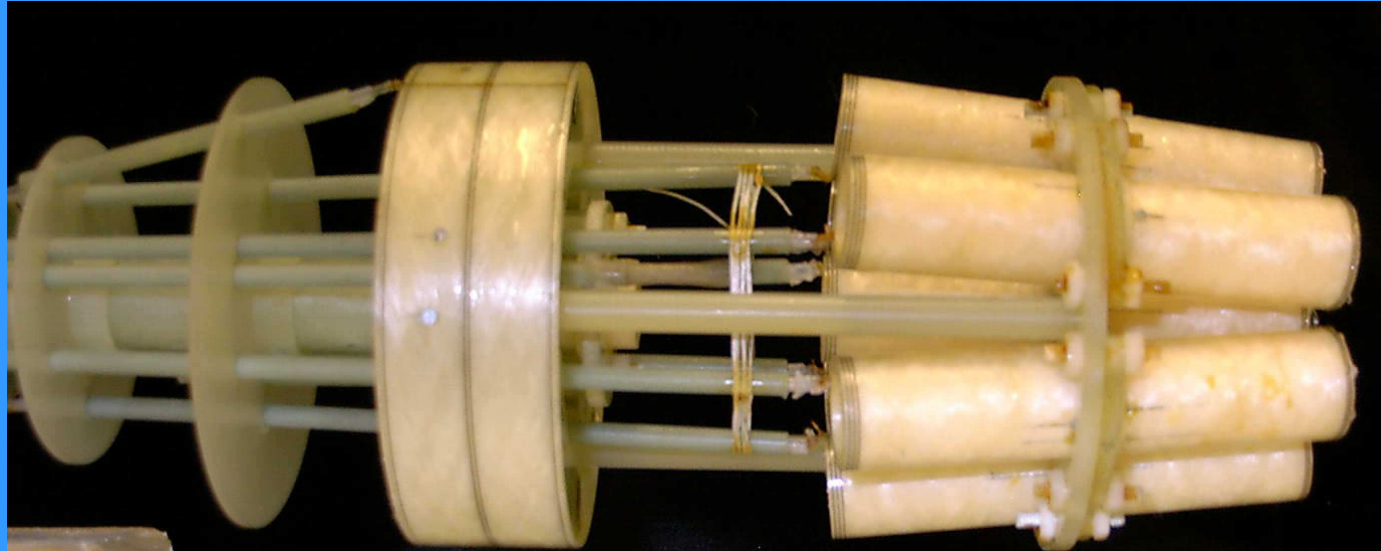


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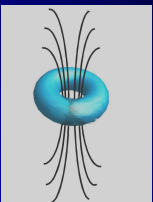


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# Second-order gradiometer Sensor Array



Measures 2<sup>nd</sup> derivative of magnetic field to minimize background pickup of external fields. Permits operation without the need for a magnetically shielded room.



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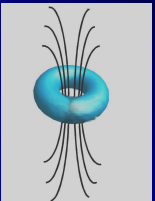


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# Methodology

## Procedure for Measuring Remanence Fields

- 1) Antibody-nanoparticles injected into subject ( $< 1\text{mg Fe}$ ).
- 2) Subject placed under sensor system
- 3) Magnetizing pulse applied (38 Gauss applied for 0.30 sec)
- 4) Remanence fields measured for two Seconds
- 5) Magnetic moment and location of source(s) obtained
- 6) Number of detected cells determined



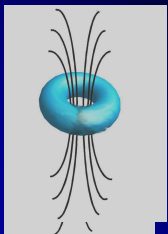
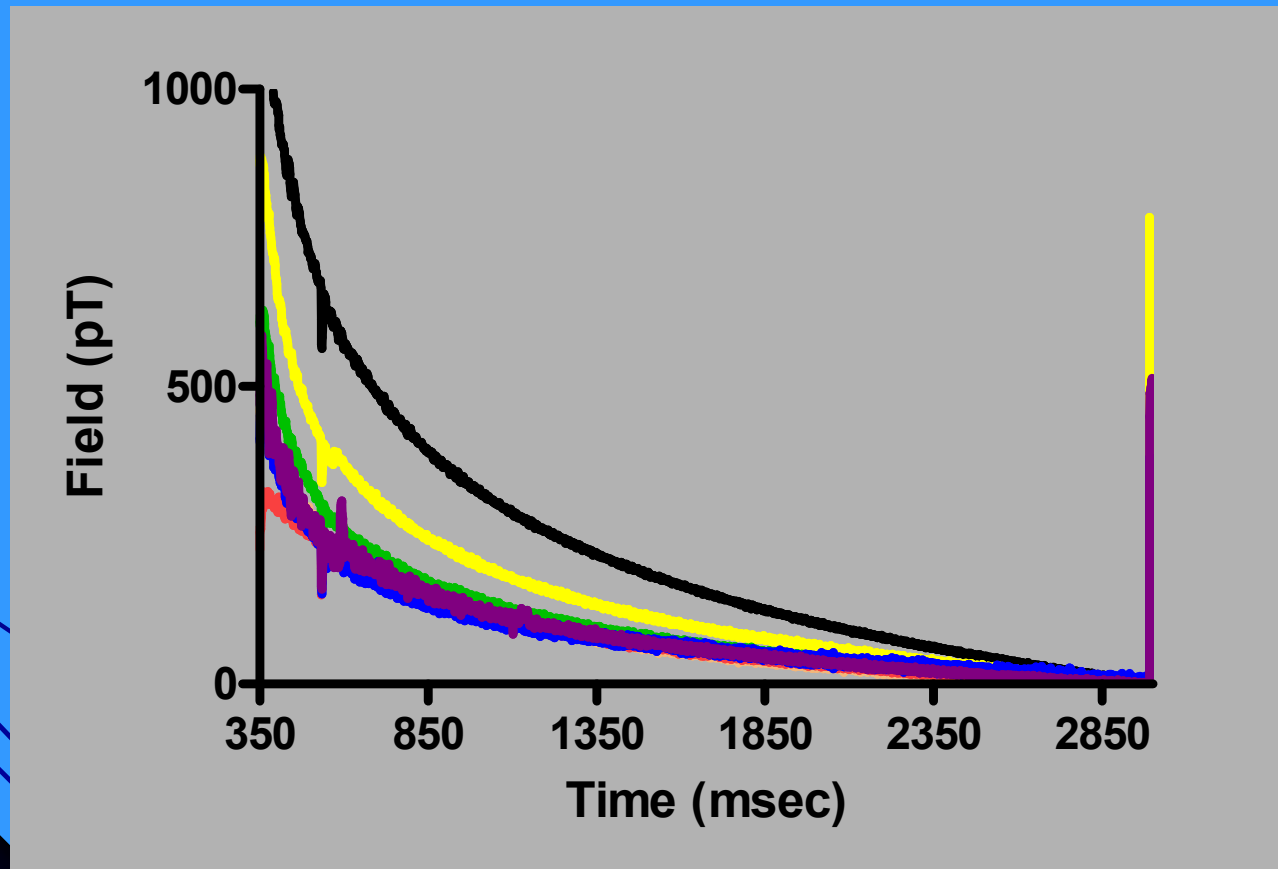
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E R Flynn and H C Bryant, "A biomagnetic system for *in vivo* cancer imaging," *Physics in Medicine and Biology* 50 (2005) 1273-1293



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# Example of 7-channel SQUID remanence fields



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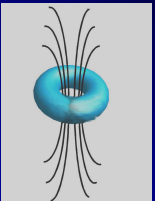


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# Disease detection procedure

## Calibrate Cell Sensitivity:

- Measure magnetic moment per particle by fitting magnetization curve to Langevin function
- Measure magnetic moment of cell sample with known number of cells
- Calculate number of nanoparticles/cell for each cell type



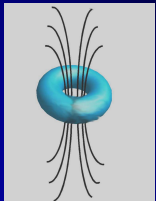
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# Superparamagnetic Particles and the Detection and Imaging of Disease

## Detecting and Locating Cancer

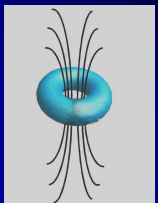
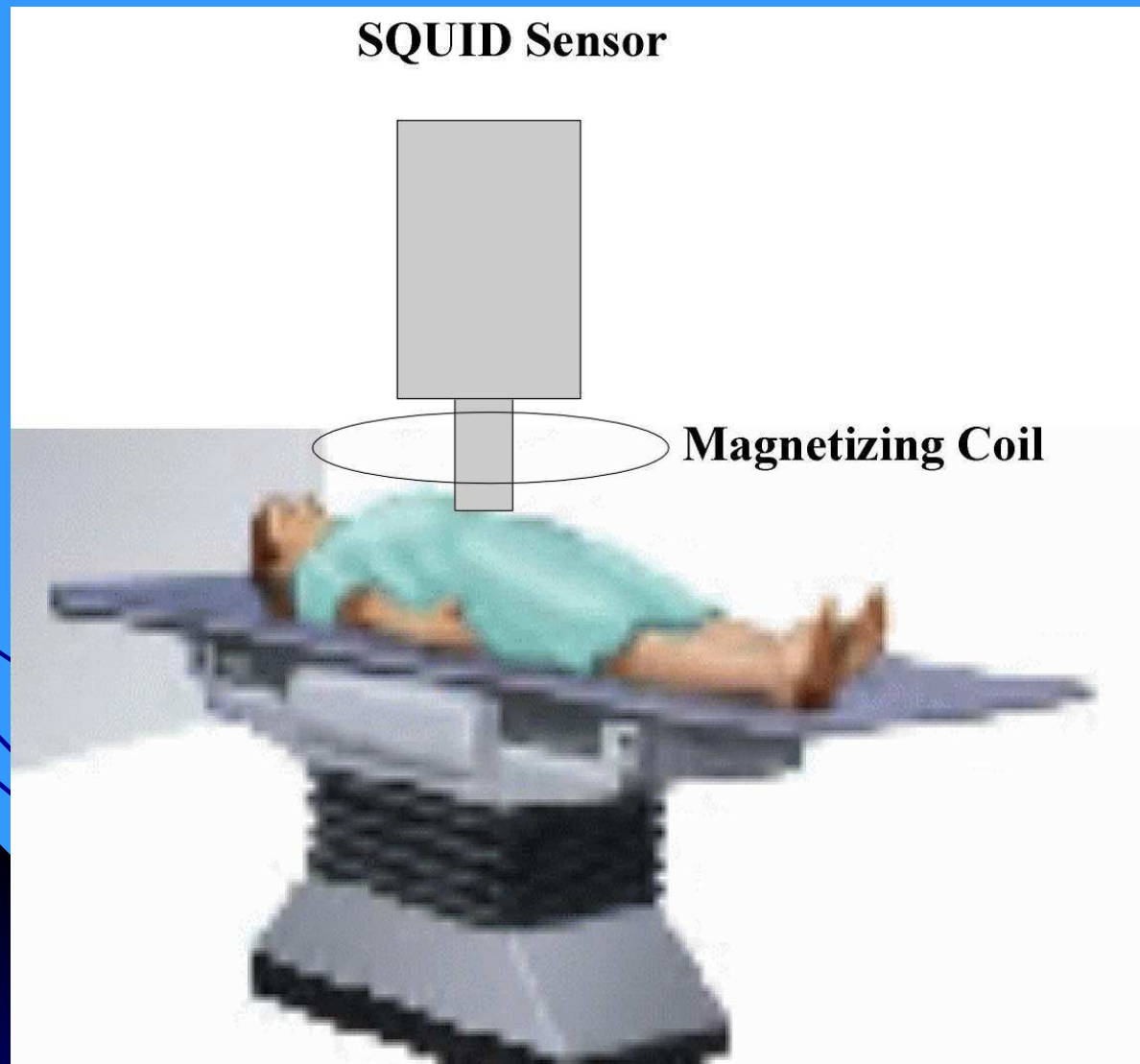


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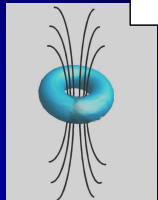
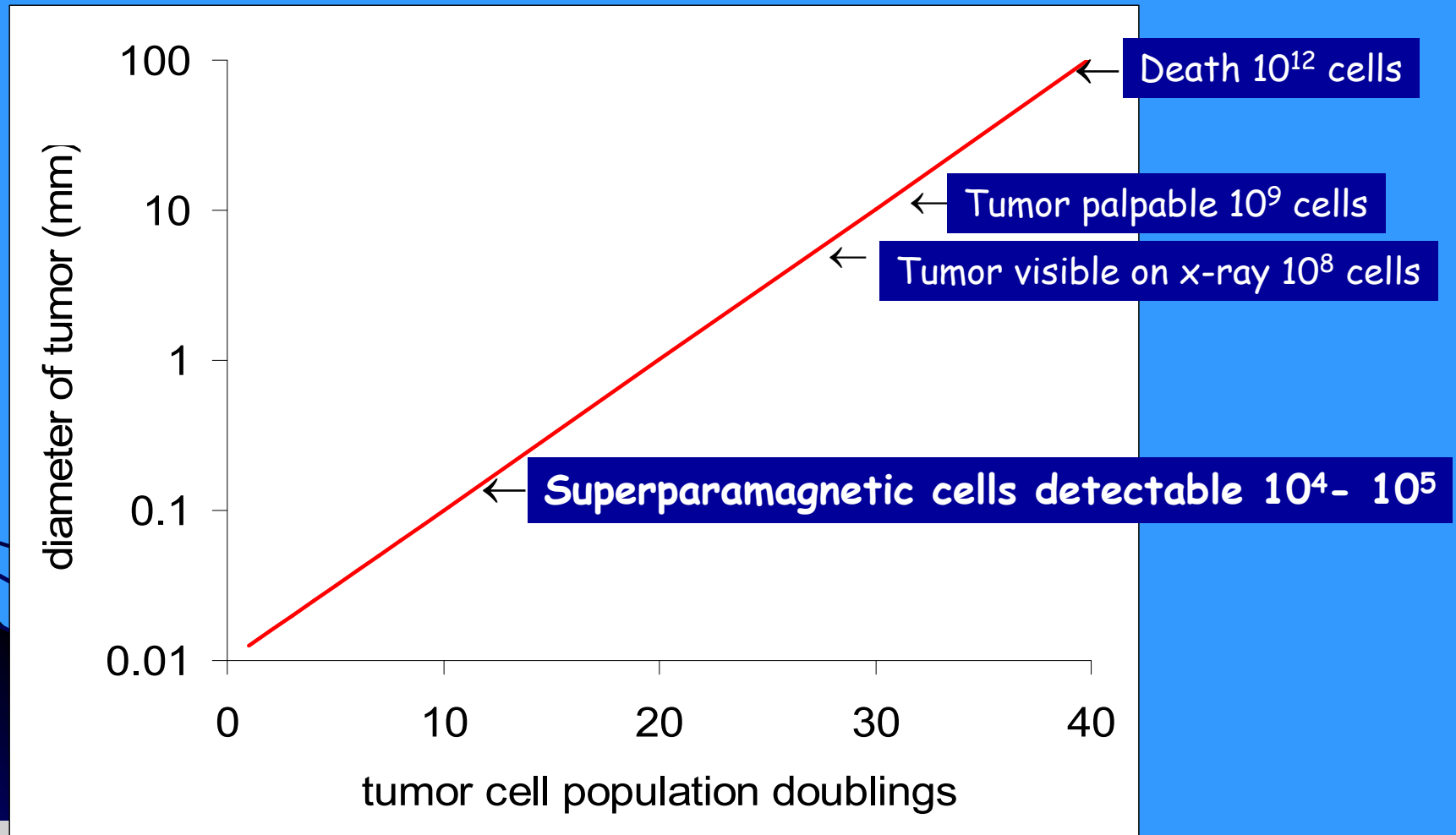
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# Clinical Arrangement in unshielded environment





# Growth of human tumor



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Reference: *The Cell* p. 1316



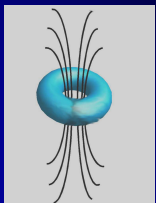
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# Breast Cancer

Breast Phantom  
with two vials  
of live breast  
Cancer cells  
(MCF-7)  
Coupled to  
Magnetic  
nanoparticles with  
HER-2 antibodies



Sensitivity for breast cancer  
cells =  $10^5$  cells for depths up to  
8 cm into breast. Imaging  
accuracy is  $\pm 3$  mm.



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# Breast Cancer Markers and Cell Lines currently under study

## Antibodies:

HER-2 antibodies

CA27.29 Breast Tumor Marker present on epithelial cells and elevated in breast cancer (33% in early and 67% in late stage cancer)

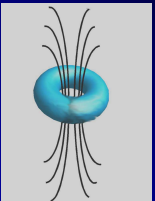
CA15-3 also elevated but not as specific

## Angiogenesis:

*VIP01 (vascular imaging peptide 01)* binds to the integrin  $\alpha_v\beta_3$  shown to be overexpressed at sites of neovascularization and metastasis.

## Cell Lines Available:

BT-549, MDA-MB-436, MDA-MB-134-VI, HCC202, HCC1008, ZR-75-1, T-47D, MDA-MB-231, and BT-474.



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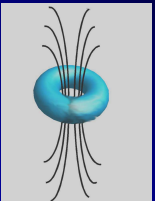
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# Mouse Model of Human breast cancer

SCID Mouse with  
human breast  
cancer Xenograft  
on flank.

MCF7 Cancer Cells

Mouse injected  
with Nanoparticles  
coupled to HER-2  
antibodies



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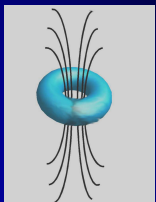
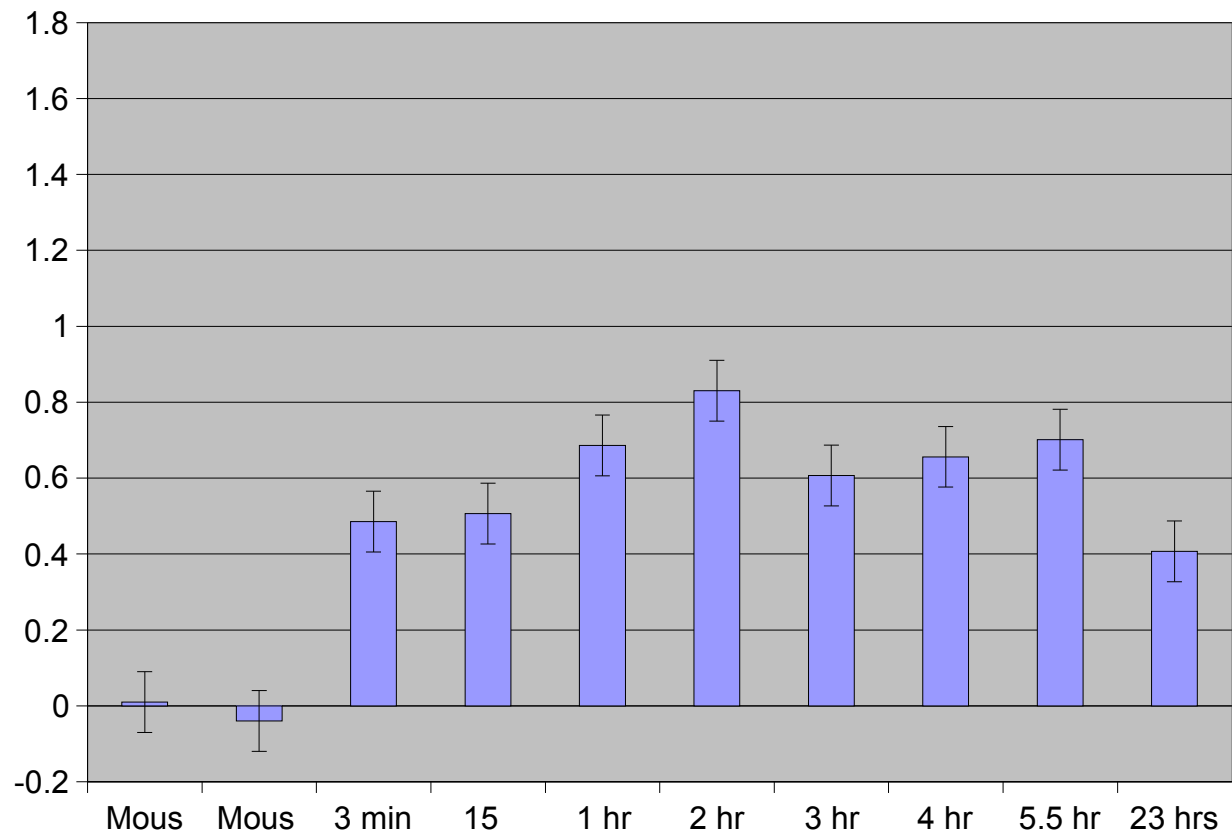
# Results of Mouse Tumor Study

Magnetic moment  
( $\times 1\text{E}+07 \text{ A}\cdot\text{m}^2$ )

as a function of  
time from mouse  
tumor after  
injection of  
nanoparticles.

$\mu = 1.0 = 2.6 \times 10^{10}$   
nanoparticles =  
 $\sim 3 \times 10^6$  cancer  
cells  
( $\sim 1 \times 10^4$  np/cell)

Magnetic Signal from Mouse after Intratumoral injection



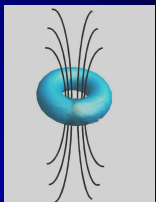
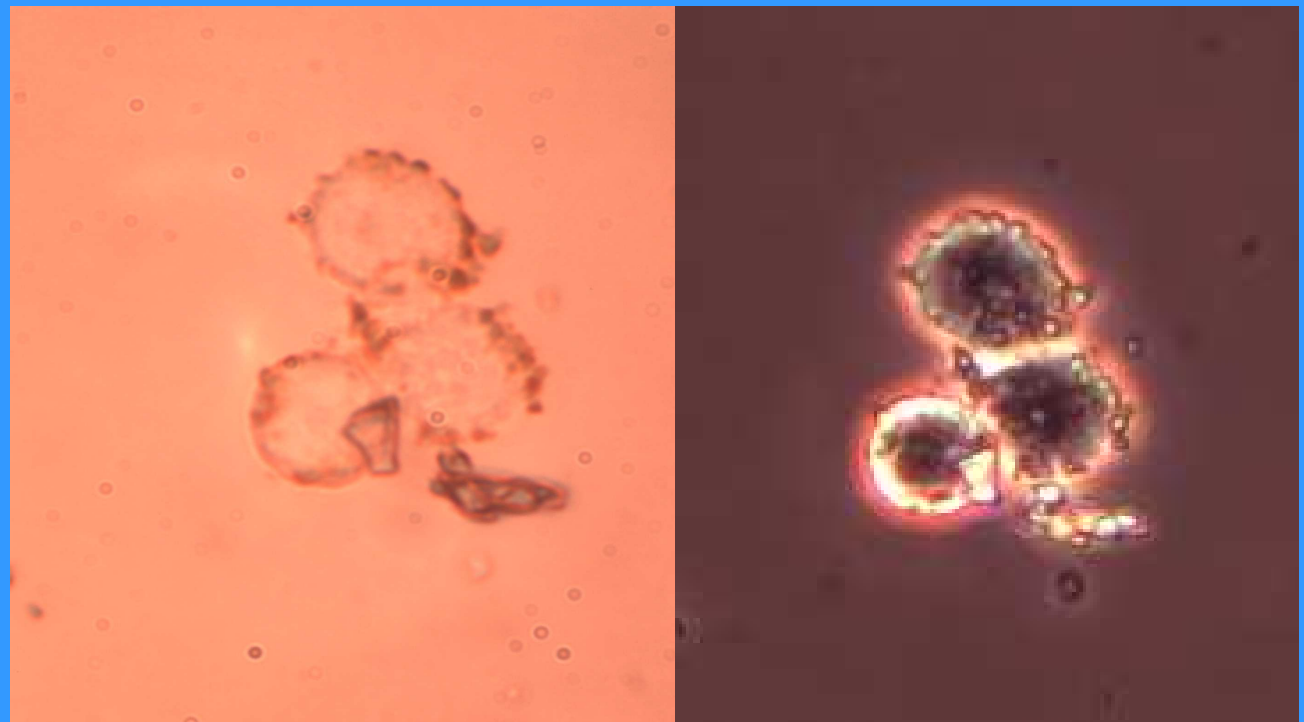
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# Labeling of Breast Cancer Cells with magnetic NP

BT474 breast tumor cells, labeled with SiMag 1411 Carboxyl magnetic NP, coupled to anti-her2 Ab. Left is bright-field image of BT474 cells showing NP bound to surface, Right image is dark-field image.



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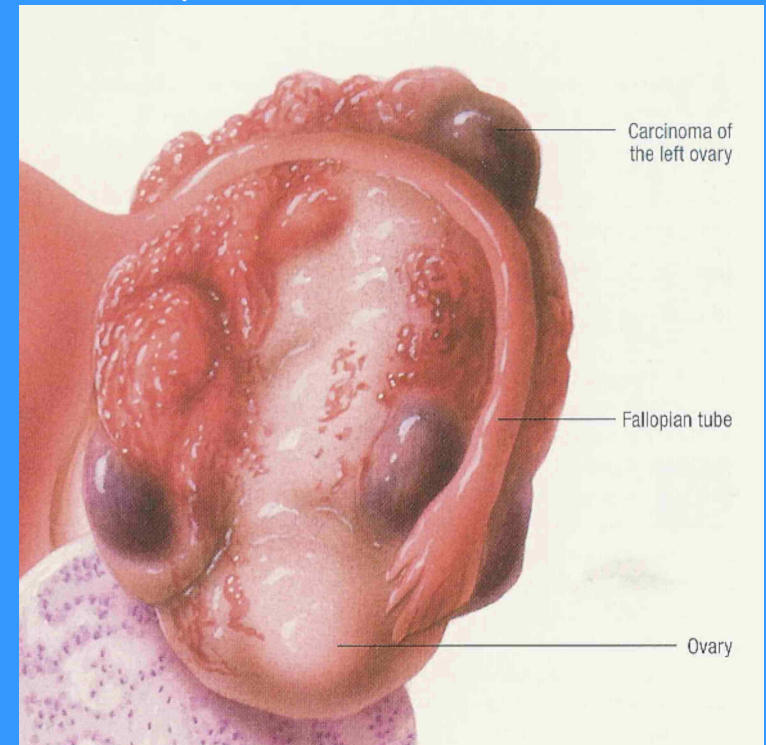


# Ovarian Cancer

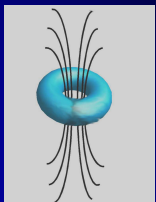
Anatomical Model under sensor system.



Ovary with Carcinomas



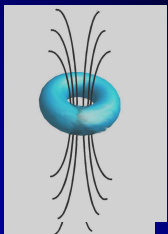
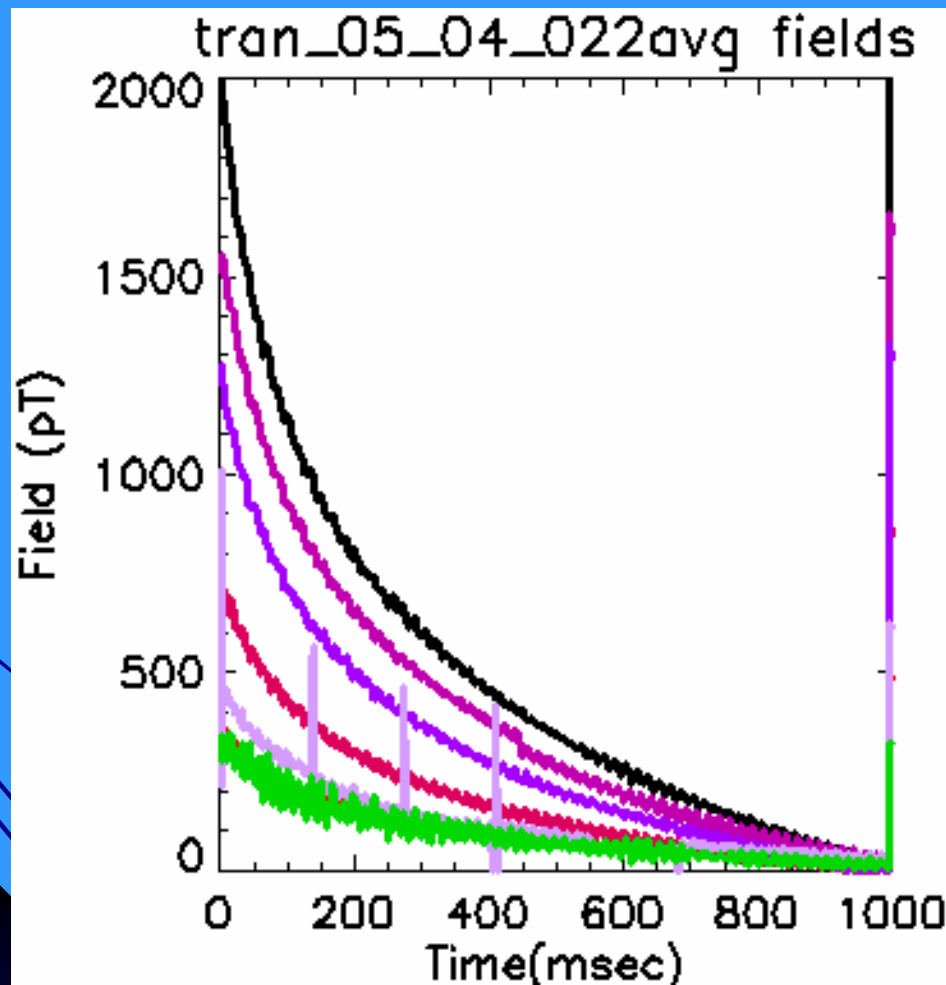
Nanoparticle in-vivo imaging permits several consecutive injections of different markers to improve sensitivity and specificity; e.g., CA125 & HMFG1/G2 give 95% sensitivity 93% specificity



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# 7-channel SQUID remanence fields from 500,000 live ovarian cells coupled to nanoparticles with CA-125 markers.



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# Live Ovarian Cell lines

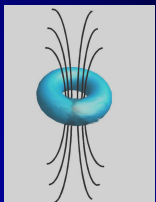
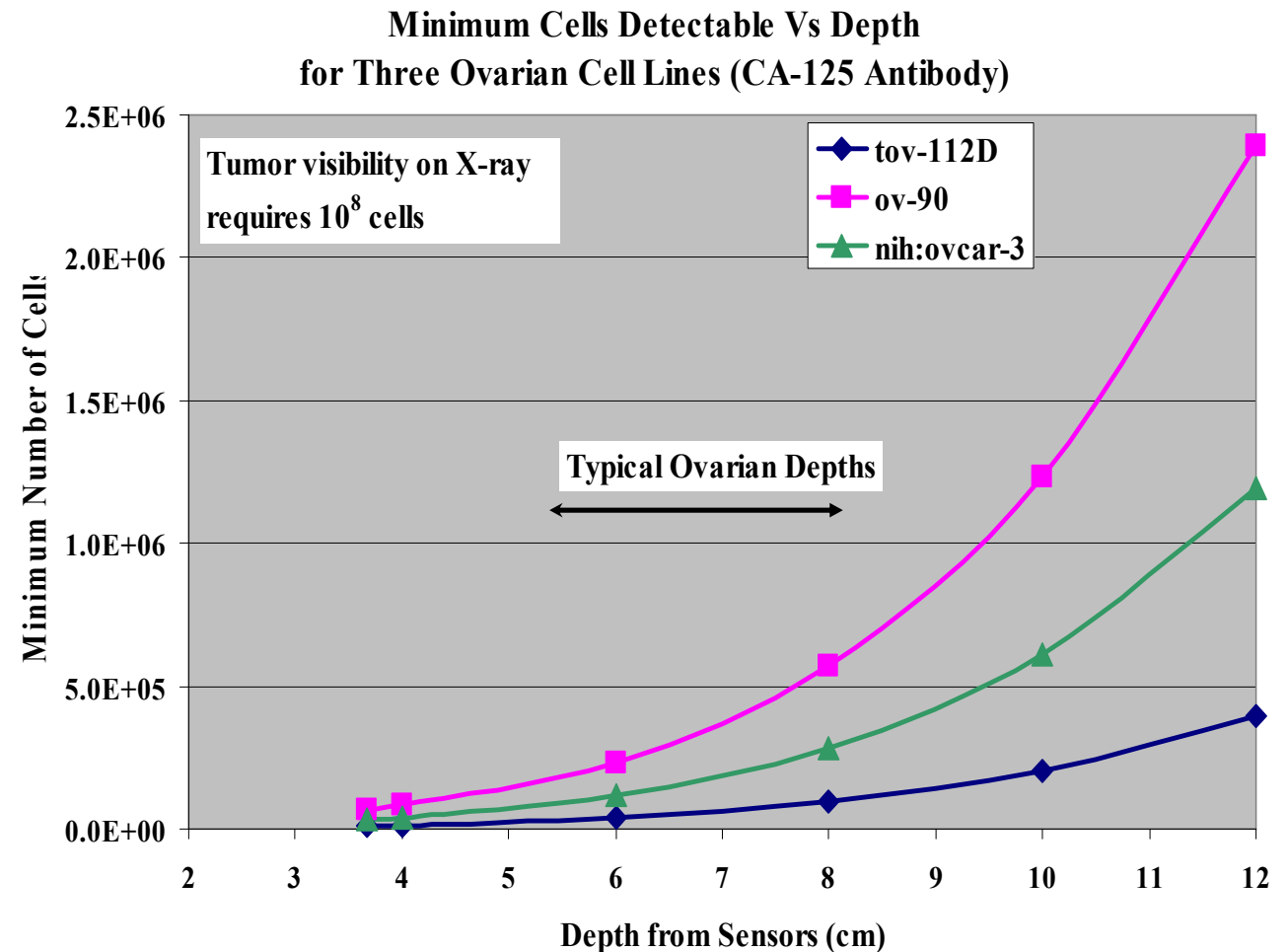
Marker: CA125

Cell Line NP/cell

tov-112D  $2.04 \times 10^4$

ov-90  $3.35 \times 10^3$

ovcar-3  $6.73 \times 10^3$



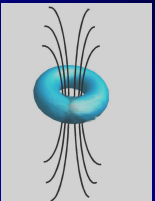
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# Superparamagnetic Particles and the Detection and Imaging of Disease

## Detecting Rejection of Transplanted Organ



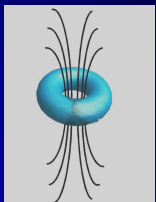
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# In-Vivo Detection of the Rejection of a Transplanted Organ

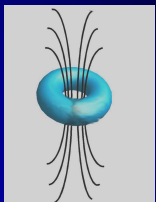
- 1) Rejection of a transplanted organ occurs by T-cells that attach to foreign Human-Leukocyte-Associated antigens on the donor cells and kill them.
- 2) T-cells accumulate in small nodules within the organ.
- 3) T-cells can be targeted with specific antibodies, conjugated to magnetic nanoparticles, and detected in-vivo by SQUID sensors.
- 4) Method minimizes the need for painful biopsies.
- 5) Monitoring for T-cell presence will be used to determine the amount of anti-immune system drugs being administered.



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# Each T-cell will have $5 \times 10^4$ CD3 Antibodies conjugated to nanoparticles

- 1) Current SQUID system detects  $10^5$  cells at 4 cm and  $10^6$  at 8cm.
- 2) This corresponds to the amount in ~100 micron diameter nodules.
- 3) Completely rejected organ may contain  $10^{10} - 10^{11}$  T-cells.
- 4) Transplanted organs include kidney, heart, liver and lung.

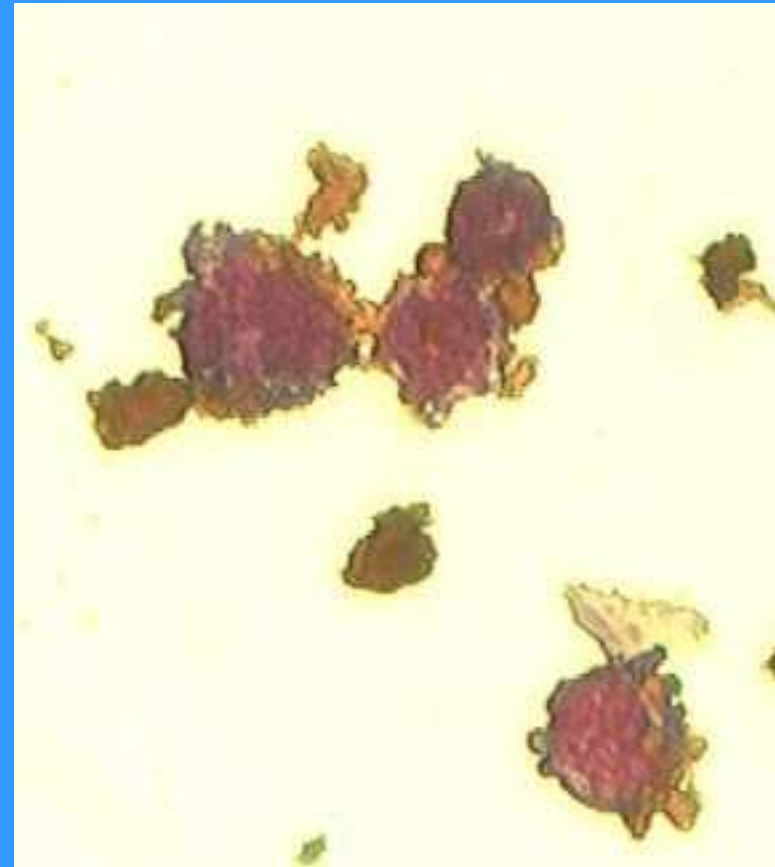


# Specific Binding of Nanoparticles to Cells

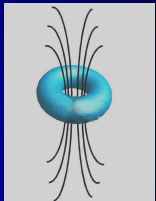
Jurkat T-Cells CD2 Antibody  
(non-specific)



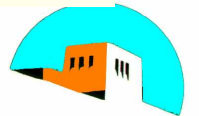
Jurkat T-Cells +CD3 Antibody  
(specific)



Jurkat cells are leukemia T-cells



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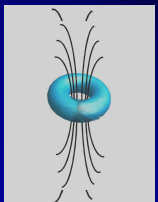
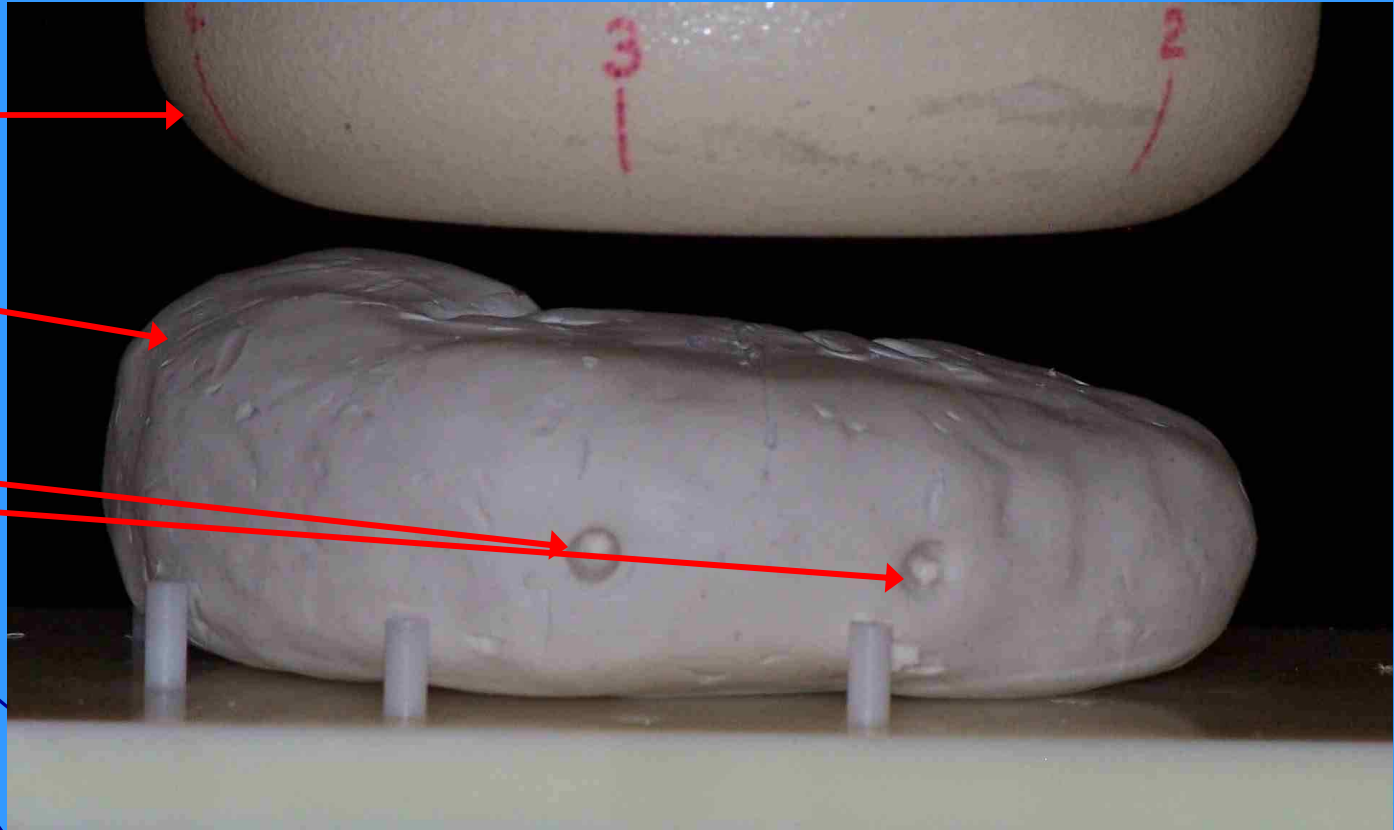
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# Kidney Phantom containing two sources of Live Jurkat T-cells under SQUID sensor

Sensor

Kidney

T-cells



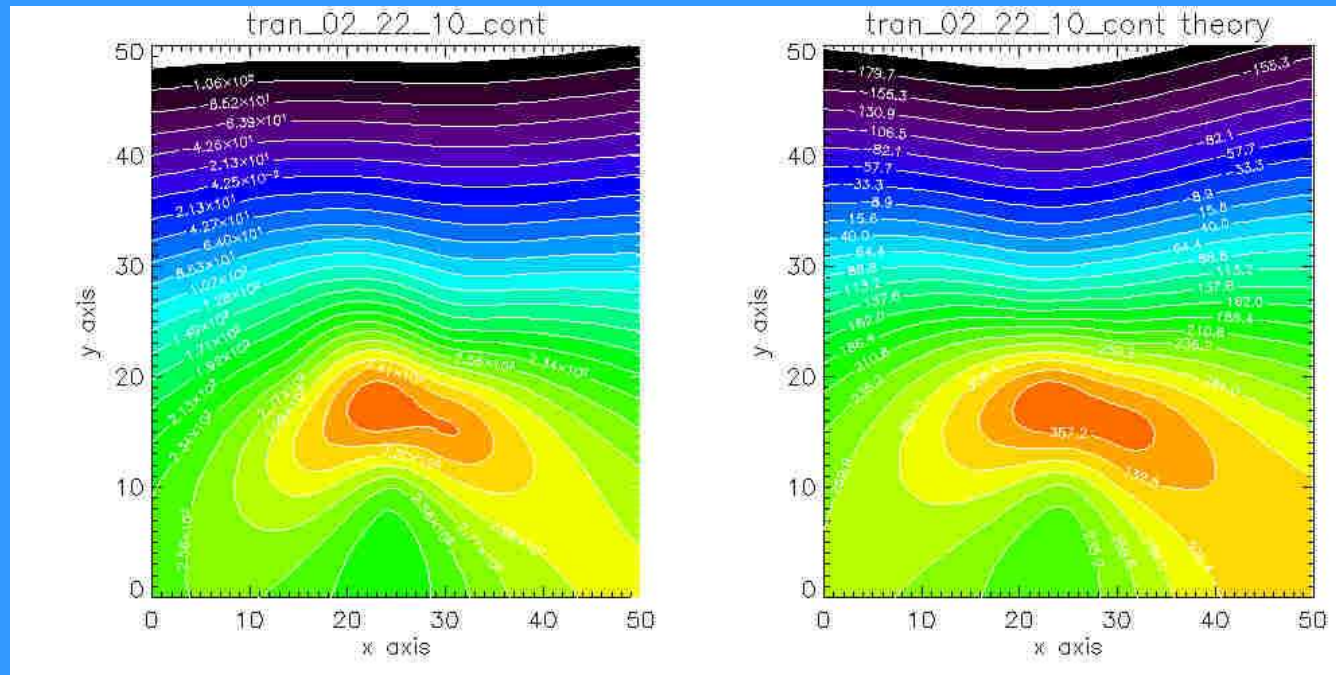
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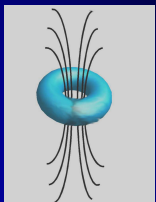
# Magnetic Field Contours of Data and Theory for two Live Cell Sources in Kidney Phantom



**Data Contours**

**Theory Contours**

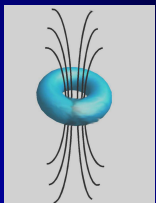
Source 1	x	y	z	m(A-m <sup>2</sup> )	Cells
measured	1.4±.3	-1.1±.3	5.5±.3	1.52E-07	4.3E+06
imaged	1.3±.4	-1.9±.4	5.0±.3	1.45E-07	4.1E+06
Source 2	x	y	z	m(A-m <sup>2</sup> )	Cells
measured	-2.7±.3	-1.5±.3	5.5±.3	1.60E-07	4.6E+06
imaged	-2.7±.4	-2.2±.3	5.0±.3	1.60E-07	4.6E+06



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# Superparamagnetic Particles and the Detection and Imaging of Disease

## Diseases of the Brain



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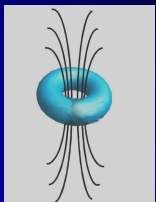
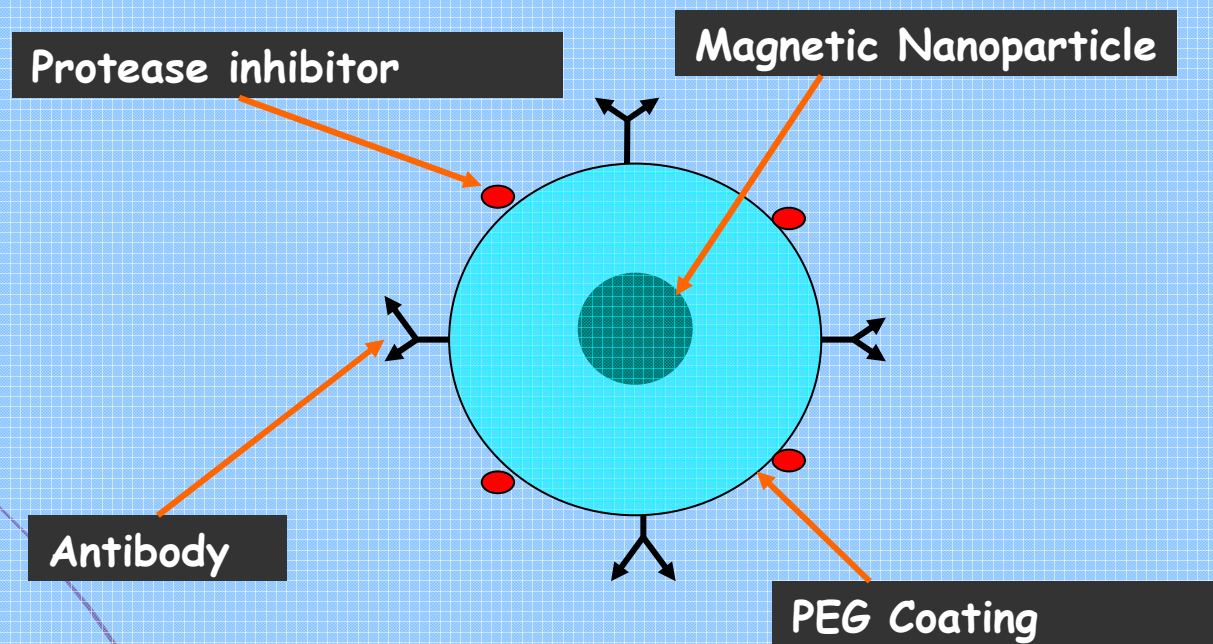


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# Diseases of the Brain

Nanoparticles and SQUID detection can be used to detect, and potentially treat, disease's of the brain; e.g., Alzheimer's and Multiple Sclerosis.

Specific coatings such as PEG help to get past the BBB with magnetic nanoparticles carrying Antibodies and Drugs



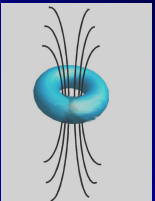
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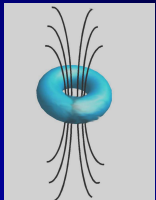
# Methodology of Detection and Treatment of AD

- 1) Use nanoparticles coated with PEG to get by BBB
- 2) Use antibodies for Amyloid Plaque and Tau
- 3) Inject into patient and detect and localize plaque and Tau deposits
- 4) Determine presence and state of AD
- 5) Inject nanoparticles with antibodies and anti-plaque drug
- 6) Magnetically concentrate particles over localized plaque sites



# Measuring Sources in the Brain

Model of brain placed in skull with multiple diffuse nanoparticle sources to measure multiple extended sources



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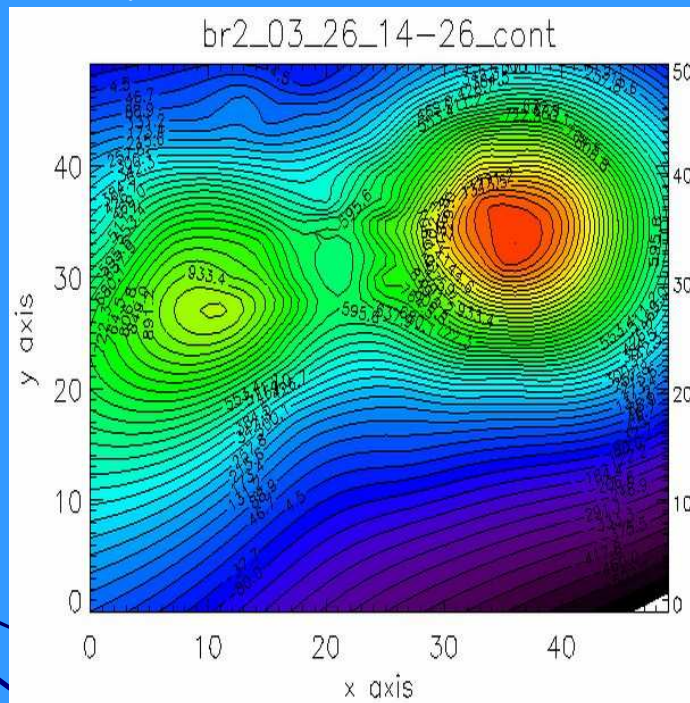


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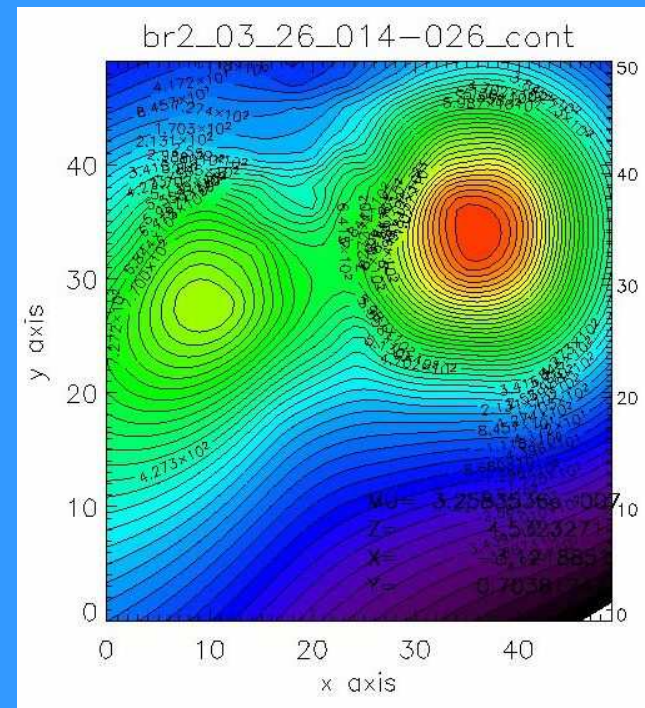


# Localization of Nanoparticle Sources in the Brain

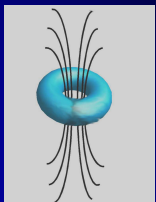
## Experimental Fields



## Theoretical Fields



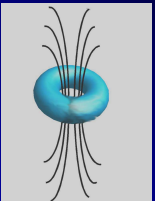
Dipole1:	X	Y	Z	M	N
	-3.12	0.70	4.53	3.26	
	7.2E+10				
Dipole2:	X	Y	Z	M	
	2.19	1.85	4.31	4.07	



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# Alzheimer Disease Status

- 1) Primary Imaging analysis completed
- 2) Antibodies for Amyloid Plaque and Tau
- 3) Antibodies coupled to Nanoparticles
- 4) Flash-frozen brain slices of AD Patients
- 5) AD Mouse model under development  
(UMN have developed AD mouse)

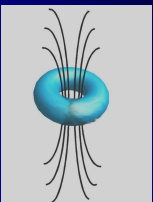




# Summary

- (1) SQUID sensor sensitivity can detect and image brain magnetic fields and targeted nanoparticles in disease.
- (2) Measurement of natural biomagnetic fields can be used to understand brain function
- (3) Measurement of natural biomagnetic fields can be used to understand the working mind
- (4) Magnetic nanoparticles and weak field sensors can be used for early disease detection.
- (5) Nanoparticle applications include cancer, leukemia, transplant rejection, and brain diseases.
- (6) Treatment options include multi-function nanoparticles for localization and magnetic concentration or hyperthermia.

This research was funded in part by:  
The National Cancer Institute  
The National Institute for Allergies and Infectious Disease  
The US Department of Defense



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# Primary Participants

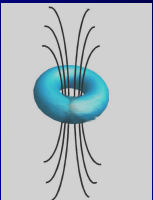
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H. C. Bryant<sup>1,3</sup>,  
D. A. Sergatskov<sup>1,4</sup>,  
R. S. Larson<sup>3</sup>, D. Lovato<sup>3</sup>, L. Sillerud<sup>3</sup>, J. Jaetao<sup>3</sup>  
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D. Huber<sup>6</sup>  
A. Georgopoulos<sup>7</sup>, A. Leuthold<sup>7</sup>

<sup>1</sup>Senior Scientific, LLC, <sup>2</sup>New Mexico Resonance

<sup>3</sup>University of New Mexico, <sup>4</sup>FermiLab,

<sup>5</sup>Chemicell GmbH, Germany, <sup>6</sup>CINT-Sandia National Lab,

<sup>7</sup>VA Hospital and UMN, Minneapolis, MN



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